

Cyclopamine Decreased the Expression of Sonic Hedgehog and its Downstream Genes in Colon Cancer Stem Cells

BAT-ERDENE BATSAIKHAN, KOZO YOSHIKAWA, NOBUHIRO KURITA, TAKASHI IWATA,
CHIE TAKASU, HIDEYA KASHIHARA and MITSUO SHIMADA

Department of Surgery, Institute of Health Biosciences, The University of Tokushima, Tokushima, Japan

Abstract. *Background:* Most solid cancers including colon cancer are believed to be initiated from and maintained by cancer stem cells (CSCs), that are responsible for treatment resistance, resulting in tumor relapse. The aim of this study was to clarify the possible role of the Sonic Hedgehog (Shh) signaling pathway in the regulation of cancer stem cells. *Materials and Methods:* The HCT-116 cell line was cultured with fetal bovine serum in RPMI-1640 medium and its sphere was grown in serum-free non-adherent culture. Gene expressions were analyzed by quantitative real-time polymerase chain reaction (qRT-PCR) from cells treated with and without cyclopamine. *Results:* HCT-116 sphere-derived cells grown in serum-free, non-adherent culture, showed significantly increased expression of stem cell markers, Shh downstream genes and epithelial-mesenchymal transition (EMT) markers compared to parental cells grown in conventional culture. The expression of stemness markers, Shh downstream genes and EMT markers were higher in cancer spheres than the parental cell line and down-regulated by cyclopamine treatment in a dose-dependent manner. *Conclusion:* Overall, these findings show that cyclopamine treatment could down-regulate the expression of stemness markers, shh downstream genes and EMT markers on HCT-116 spheres.

Cancer stem cells (CSCs) play an important role in cancer development, because of their main characteristics, which are their self-renewing capacity, chemoresistance, and their tumorigenic capacity (1). They may also play a crucial role

Correspondence to: Kozo Yoshikawa, MD, Ph.D., FACS, Department of Surgery, Institute of Health Biosciences, The University of Tokushima, 3-18-15 Kuramoto-cho, Tokushima 770-8503, Japan. Tel: +81 886337137, Fax: +81 886319698, e-mail: yoshikawa.kozo@tokushima-u.ac.jp

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in the initiation, progression and recurrence of cancer. Several therapeutic strategies have been suggested to target CSCs. Inhibiting the key signaling pathways that are active in CSCs is one of the most promising strategies for treatment of cancer. Hedgehog signaling (Shh) pathways are essential to regulate the self-renewal of CSCs and are aberrantly activated in a variety of cancers.

The classic inhibitor is cyclopamine, a naturally-occurring teratogenic alkaloid. It disrupts cholesterol bio-synthesis and specifically antagonizes the Shh signaling pathway through direct interaction with Smoothed (SMO), which functions upstream of *GLI1* (2).

Epithelial-mesenchymal transition (EMT) is a process in which cells lose their epithelial character and acquire a migratory mesenchymal phenotype (3). EMT is induced by transforming growth factor- β 1 (TGF- β 1), that is involved in the promotion of tumor invasion and metastasis. It is also closely-related to drug resistance of tumor cells (4). Loss of E-Cadherin, which is a main determinant of epithelial tissue organization and cell polarity, is considered a hallmark of EMT (5). Recent studies show a repression of E-Cadherin related to enhanced invasiveness and metastasis, while conversely, up-regulation of E-Cadherin decreases in tumor malignancy (6, 7).

To eradicate colon cancer and prevent recurrence, it is imperative that HCT-116 CSCs should be specifically and efficiently inhibited. In addition, CSCs undergoing metastasis often express epithelial-to-mesenchymal transition and the high EMT markers have been observed in dissemination of carcinoma cells from primary epithelial tumors. The interaction between stemness characteristics and EMT expression has advanced results in recent studies. Researchers have shown that EMT acquires CSC properties, increases cancer cell tumorigenicity and shows a crucial link to metastasis.

The cancer-preventive effects of cyclopamine are widely supported by results from epidemiological, cell culture, animal and clinical studies. This property has been mainly attributed to the capacity in inhibiting cancer growth in

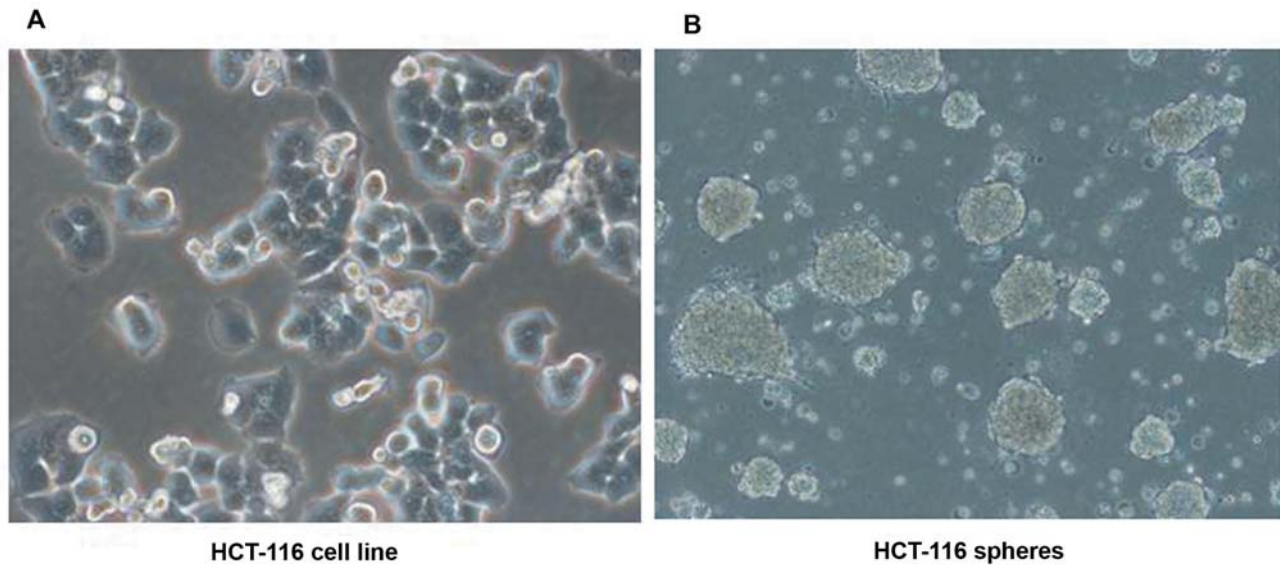


Figure 1. Representative images of HCT-116 colon cancer cell line. A) HCT-116 colon cancer cell line B) HCT-116 spheres.

various cancers by several mechanisms. CSCs are believed to be more drug-resistant compared to parental cells and so can be ultimately inducing tumour recurrence after the completion of treatment. Thus, removal of CSCs becomes more and more crucial to chemotherapy and drugs that selectively target CSCs offer a greater promise for cancer treatment. The aim of this study was to determine the mRNA expression levels of stemness markers, Shh downstream genes and EMT markers on HCT-116 cancer sphere and to explore the possible role of cyclopamine on the expression of these genes.

The goal of this study was to clarify the possible role of Shh signaling pathway's genes in the regulation of CSCs-, and effect of cyclopamine to EMT, tight junction proteins, and CSCs markers.

Materials and Methods

Cells and culture conditions. The human colon cancer (HCT-116) cell line was bought from the American Type Culture Collection (ATCC, 10801 University Boulevard Manassas, VA, USA). The cell line was maintained as a monolayer in Roswell Park Memorial Institute medium 1640 (RPMI 1640) (Invitrogen, Gibco, Carlsbad, CA, USA) supplemented with 10% fetal bovine serum (FBS) (Invitrogen/Gibco-), 100 IU/ml penicillin G and 100 µg/ml streptomycin at 37°C in a humidified 5% CO₂ incubator. The cells were collected and washed to remove serum, then suspended in serum-free Dulbecco modified Eagle's minimal essential medium (DMEM/F12) (Invitrogen/Gibco-) supplemented with 100 IU/ml penicillin, 100 µg/ml streptomycin, 20 ng/ml human recombinant epithelial growth factor (hrEGF) (BD Biosciences, cat.n: 354052, San Diego, CA, USA), 10 ng/ml human recombinant basic

fibroblast growth factor (hrbFGF) (BD Biosciences, cat.n: 354060-), 2% B27 supplement without vitamin A, 1% N2 supplement (Invitrogen).

Isolation of RNA. Total RNA was extracted from HCT-116 parental cells and its sphere-, using TRIzol reagent (Invitrogen). cDNA was prepared using the high capacity cDNA Reverse Transcription Kit (Applied Biosystems, Foster, CA, USA), following the manufacturer's instructions. The RNA pellets were stored at -80°C until use.

Quantitative reverse transcriptase polymerase chain reaction (qRT-PCR). QRT-PCR was performed with SYBR Green master mix real-time core reagents on an ABI 7500 (Applied Biosystems-) according to the manufacturer's instructions. Primers for qRT-PCR were as follows: Taqman® stemness genes: Oct-4 (*POU5F1*) Hs03005111_g1; *NANOG* Hs02387400_g1; CD-44 Hs01081473_m1; *EpCAM* Hs00901885_m1. Shh pathway downstream genes: *PTCH1* Hs00181117_m1; *SMO* Hs01090242_m1; and Shh downstream gene *Gli1* Hs01110766_m1. Tight junctions are Claudin-4 (*CLDN4*) (Hs00533616_s1) and Occludin (*OCLN*) Hs00170162_m1.

Cyclopamine treatment. The HCT-116 cell line was plated in 10 cm dishes and incubated for 48 h at 37°C, in 5% CO₂ and 95% humidity. Then the cells were transferred into sphere forming medium and incubated for 12 days at 37°C, in 5% CO₂ and 95% air. The sphere cells were exposed to cyclopamine at 2 µM and 5 µM or no cyclopamine as a control group for 48 h. Lastly, the cells were harvested and subjected to RNA extraction.

Statistical analysis. Data were obtained from experiments performed at least 3 times with a minimum of triplicates. All values in the figures and text are shown as mean±standard deviation (SD). Statistical analyses were performed using the Stat.View (SAS

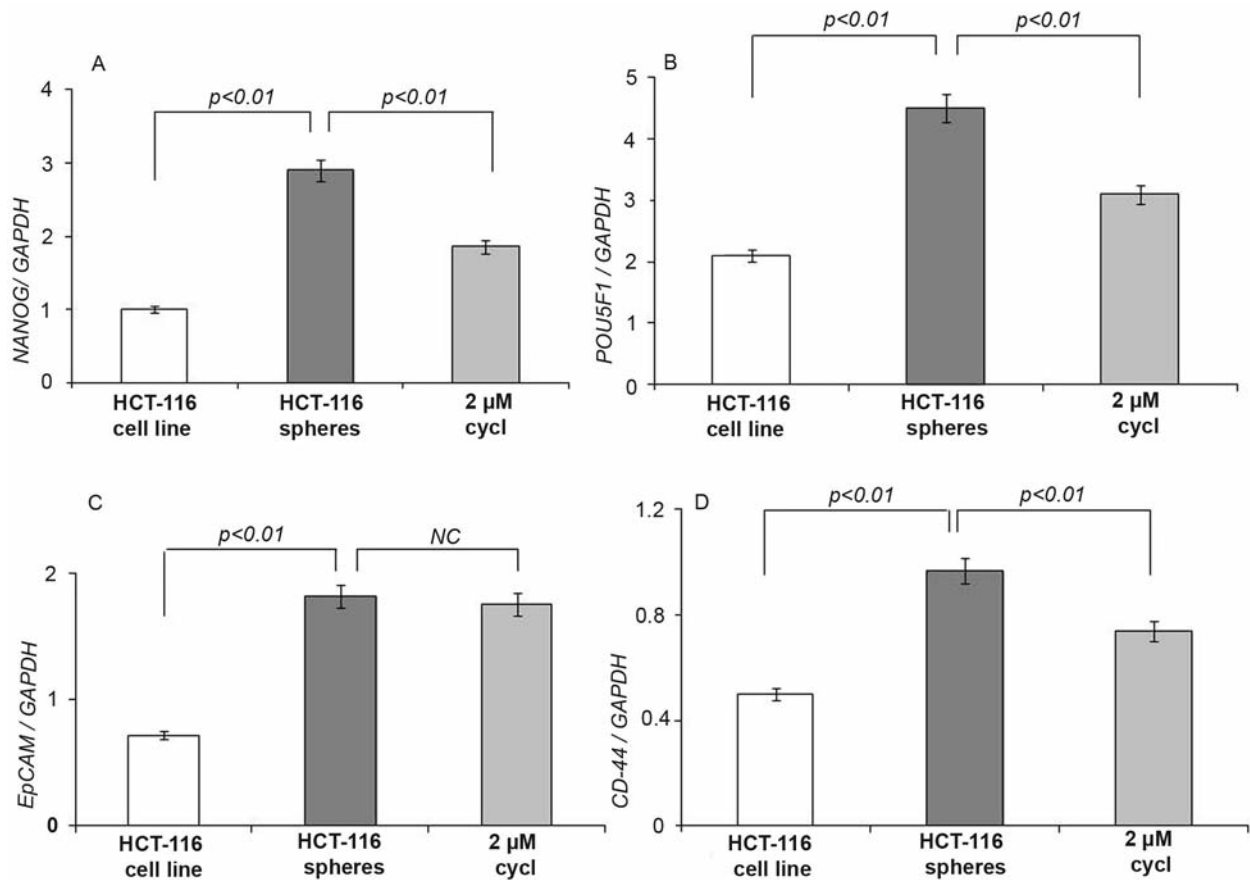


Figure 2. mRNA expression level stemness markers before and after cyclopamine treatment in HCT-116 spheres. A) The mRNA level of NANOG was significantly expressed in HCT-116 spheres compared to HCT-116 cell line. After cyclopamine treatment the mRNA was down-regulated in HCT-116 spheres. B) The mRNA level of POU5F1 was significantly expressed in HCT-116 spheres compared to HCT-116 cell line. After cyclopamine treatment the mRNA level was down-regulated in HCT-116 spheres. C) The mRNA level of EpCAM was significantly expressed in HCT-116 spheres compared to HCT-116 cell line. After cyclopamine treatment the mRNA level presented no difference in HCT-116 spheres. D) The mRNA level of CD-44 was significantly more expressed in HCT-116 spheres compared to HCT-116 cell line. After cyclopamine treatment the mRNA was down-regulated in HCT-116 spheres.

Campus Drive, Cary, NC, USA). Any significant differences among mean values were evaluated by the student's *t*-test. A *p*-value of less than 0.05 was accepted as statistically significant.

Results

mRNA level of surface marker and stemness gene before and after administration of cyclopamine in HCT-116 sphere. HCT-116 sphere successfully formed from HCT-116 parental cells (Figure 1A and B). The stemness genes (*NANOG*, *POU5F1*) and surface markers (*CD-44*, *EpCAM*) were significantly expressed in HCT-116 sphere compared to HCT-116 colon cancer cell line ($p < 0.01$). After administration of cyclopamine the mRNA level of *NANOG*, *POU5F1* and *CD-44* was reduced in HCT-116 sphere ($p < 0.01$) (Figure 2 A, B, C). However, the mRNA level of *EpCAM* was not changed after administration of cyclopamine (Figure 2C).

Regulation of mRNA level of Shh downstream genes by cyclopamine in HCT-116 sphere. As indicated on Figure 3 (A-C), a significant mRNA expression level of *PTCH1*, *SMO* and *GLII* was observed between HCT-116 sphere and its cell line ($p < 0.01$). After treatment of cyclopamine the mRNA level of these genes was significantly reduced.

mRNA expression level of the epithelial marker E-Cadherin and mesenchymal marker Vimentin, before and after cyclopamine treatment. As shown in Figure 4 A and B, the mRNA level of *CDH1* (E-Cadherin) was down-regulated in HCT-116 sphere compared to HCT-116 cell line, while the mRNA level of *VIM* (Vimentin) was overexpressed in HCT-116 sphere than in the HCT-116 cell line. After cyclopamine treatment the mRNA level of *CDH1* (E-Cadherin) was up-regulated in HCT-116 sphere, whereas

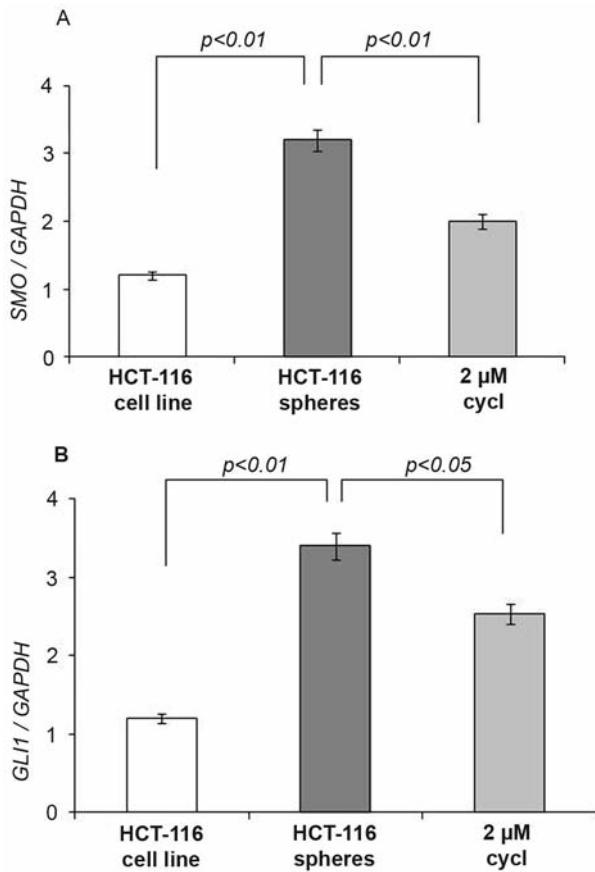


Figure 3. *Shh* inhibitor suppresses mRNA levels of *SMO* and *GLI1* in HCT-116 spheres. A) *SMO* expression before and after administration on 2 μM dose in HCT-116 spheres. B) *GLI1* expression before and after administration on 2 μM dose in HCT-116 spheres.

the mRNA level of *VIM* was down-regulated in HCT-116 sphere ($p < 0.01$).

mRNA expression level of tight junction proteins in HCT-116 sphere. The mRNA expression level of *CLDN4* (claudin-4) and *OCN* (occludin) were significantly expressed in HCT-116 sphere than the HCT-116 cell line. After administration of cyclopamine the mRNA level was down-regulated in HCT-116 sphere ($p < 0.01$).

Discussion

Colon cancer sphere of HCT-116, grown in serum-free non-adherent culture, showed increased expression of stem cell markers such as *NANOG*, *POU5F1*, *CD-44* and *EpCAM*, compared to parental cells grown in conventional cultures. Previous studies identified the expression of cell surface

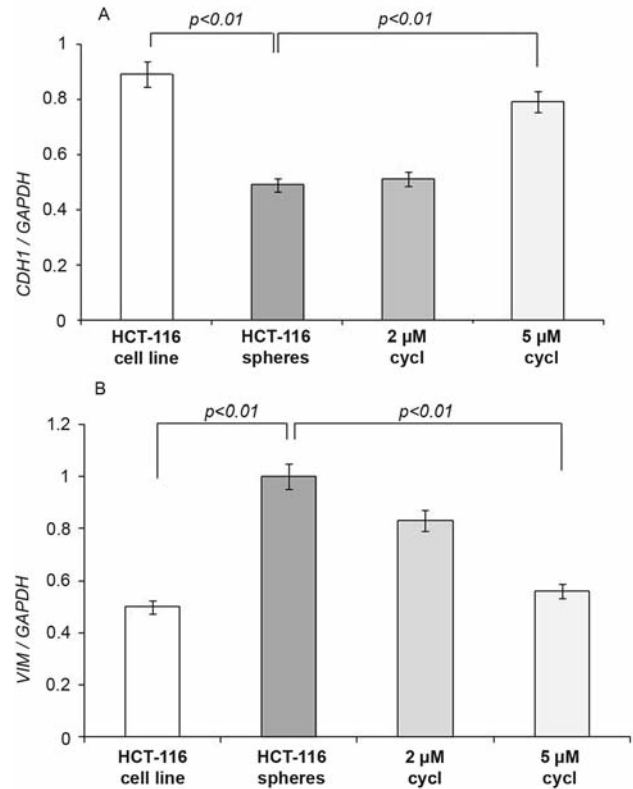


Figure 4. *Shh* inhibitor (cyclopamine) suppresses expression of EMT markers in HCT-116 spheres. A) mRNA level of *CDH1* (E-Cadherin) significantly expressed in HCT-116 cell line than HCT-116 spheres. After administration of cyclopamine on 2 μM and 5 μM dose the mRNA level was up-regulated in HCT-116 spheres. B) mRNA level of *VIM* (Vimentin) significantly expressed in HCT-116 spheres than HCT-116 cell line. After administration of cyclopamine on 2 μM and 5 μM dose the mRNA level was down-regulated in HCT-116 spheres.

markers *CD-44*, *CD24*, *ESA*, *CD-133* (8)- and *EpCAM* (9) to be highly expressed in cancer sphere. *POU5F1* and *NANOG* are essential regulators of stemness by promoting proliferative potential and self-renewal in embryonic stem cells during early embryonal development. These are specifically expressed in the human embryonic pluripotent stem cells (10). Down-regulation of stem cell markers may be a novel approach for colon cancer therapy, especially for HCT-116 CSCs. In the present study, we have convincingly demonstrated that cyclopamine treatment down-regulated the stemness markers, *NANOG*, *POU5F1* and *CD-44*, which indicates that cyclopamine could be an effective anticancer agent for HCT-116 spheres.

Several therapeutic strategies have been suggested to target CSCs. Inhibiting the key signaling pathways that are active in CSCs is one of the most promising strategies for treatment of cancer. *Shh* pathways are essential to regulate

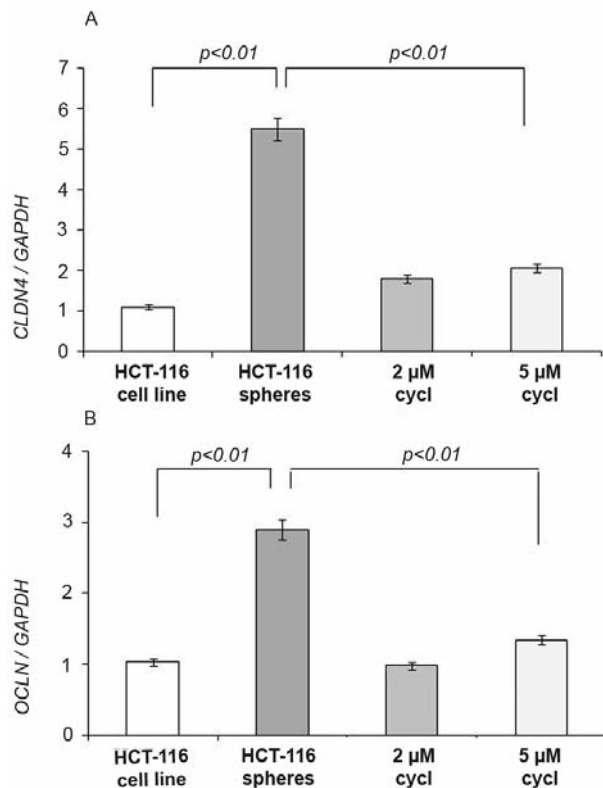


Figure 5. Cyclopamine influences in tight junction markers in HCT-116 spheres. A) mRNA level of *CLDN4* (Claudin-4) decreased after administration of cyclopamine in HCT-116 spheres. B) mRNA level of *OCLN* (Occludin) decreased after administration of cyclopamine in HCT-116 spheres.

the self-renewal of CSCs and play important role in maintaining stemness by regulating the expression of stemness genes (11). Mueller *et al.* reported a high expression of CSC markers correlated with Shh up-regulation in gemcitabine-resistant pancreatic cancer cells (12) explaining that the Shh pathway target genes could induce self-renewal, survival, and migration of cancer progenitor cells (13, 14).

Zhang *et al.* reported that the *Shh*, *PTCH1* and *GLI3* were more highly expressed in CD44⁺CD24⁺ gastric cancer stem cells, than cancer cells (15), while cyclopamine treatment was shown to inhibit up-regulation of these genes by interfering with activation of *SMO* (16, 17). In line with this, in the present study, the mRNA level of Shh downstream genes, *PTCH1* and *SMO* were significantly overexpressed in HCT-116 sphere, than the HCT-116 cell line, with cyclopamine treatment inhibiting up-regulation of these genes in HCT-116 sphere.

The Shh signaling pathway activation includes epithelial-mesenchymal transition (EMT), which is required for

migration of Shh-activated cells during tissue morphogenesis. To eradicate colon cancer and prevent recurrence, it is imperative that colon CSCs should be specifically and efficiently inhibited. In addition, CSCs undergoing metastasis often express epithelial-to-mesenchymal transition and the high EMT markers have been observed in dissemination of carcinoma cells from primary epithelial tumors. The interaction between stemness characteristics and EMT expression has advanced results in recent studies. Researchers have shown that EMT acquires CSC properties, increases cancer cell tumorigenicity and shows a crucial link to metastasis.

In the present study, the mesenchymal marker Vimentin was up-regulated in HCT-116 sphere, while epithelial marker E-Cadherin was down-regulated in HCT-116 sphere. After using cyclopamine the Vimentin was down-regulated and E-Cadherin was up-regulated in HCT-116 spheres. E-Cadherin has a down-regulated expression upon an increased expression of Shh/Gli1 (18). E-Cadherin plays a main role in the development of epithelial tissue and cell polarity (19) and its loss is a hallmark of an EMT. A previous study mentioned down-regulation of *GLI1* and showed up-regulation of E-Cadherin in gastrointestinal neuroendocrine carcinomas occurring after the use of Shh inhibitor cyclopamine (20). The results of the present study reveal that cyclopamine could influence the production of EMT by *GLI1* down-regulation, which could activate or increase mRNA level of *CHD1* (E-Cadherin) in a colon cancer sphere of the HCT-116 cell line.

In addition, tight junction proteins showed high expression of EMT genes in human breast cancer (21). Taube and Turksen reported the low or nil expression of *CLDN4* (claudin-4) that has been associated with breast cancer-derived CSCs (22, 23). However, there is a high expression of claudin-4 in ovarian CSCs (24). In our study high expression of claudin-4 and occluding in HCT-116 sphere was significantly down-regulated by cyclopamine treatment. The expression of tight junction proteins is controversial in CSCs. Future studies need to investigate the role of tight junction proteins in CSCs in depth.

In conclusion, overall, our findings show that HCT-116 cells with sphere formations possess the properties of CSCs. Using this model, we found that cyclopamine treatment down-regulated genes associated with self-renewal and invasive capacities in HCT-116 spheres. This supports the notion that cyclopamine may decrease recurrence and metastasis in colon carcinoma.

Conflicts of Interest

Bat-Erdene Batsaikhan and other co-authors declare no conflict of interest.

References

- 1 Song Z, Yue W, Wei B, Wang N, Li T, Guan L, Shi S, Zeng Q, Pei X and Chen L: Sonic Hedgehog Pathway is Essential for Maintenance of Cancer Stem-Like Cells in Human Gastric Cancer. *PLoS One* 6(3): e17687, 2011.
- 2 Low JA and de Sauvage JF: Clinical experience with Hedgehog pathway inhibitors. *J Clin Oncol* 28(36): 5321-5326, 2010.
- 3 Thiery JP, Acloque H, Huang RY and Nieto MA: Epithelial-mesenchymal transitions in development and disease. *Cell* 139(5): 871-890, 2009.
- 4 Sun XD, Liu XE and Huanq DS: Curcumin reverses the epithelial-mesenchymal transition of pancreatic cancer cells by inhibiting the Hedgehog signaling pathway. *Oncol Rep* 29(6): 2401-2407, 2013.
- 5 Yang J and Weinberg RA: Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. *Dev Cell* 14(6): 818-829, 2008.
- 6 Seidel B, Braeg S, Adler G, Wedlich D and Menke A: E- and N-cadherin differ with respect to their associated p120ctn isoforms and their ability to suppress invasive growth in pancreatic cancer cells. *Oncogene* 23(32): 5532-5542, 2004.
- 7 von Burstin J, Eser S, Paul MC, Seidler B, Brandl M, Messer M, von Werder A, Schmidt A, Mages J, Pagel P, Schnieke A, Schmid RM, Schneider G and Saur D: E-cadherin regulates metastasis of pancreatic cancer in vivo and is suppressed by a SNAIL/HDAC1/HDAC2 repressor complex. *Gastroenterology* 137(1): 361-371, 2009.
- 8 Yao J, An Y, Wie JS, Ji ZL, Lu ZP, Wu JL, Jiang KR, Chen P, Xu ZK and Miao Y: Cyclopamine reverts acquired chemoresistance and down-regulates cancer stem cell markers in pancreatic cancer cell lines. *Swiss Med Wkly* 141: w13208, 2011.
- 9 Dalerba P, Dylla SJ, Park IK, Liu R, Wang X, Cho RW, Hoey T, Gurney A, Huang EH, Simeone DM, Shelton AA, Parmiani G, Castelli C and Clarke MF: Phenotypic characterization of human colorectal cancer stem cells. *Proc Natl Acad Sci USA* 104(24): 10158-10163, 2007.
- 10 Yin X, Li YW, Zhang BH, Ren ZG, Qiu SJ, Yi Y and Fan J: Coexpression of stemness factors Oct4 and Nanog predict liver resection. *Ann Surg Oncol* 19(9): 2877-2887, 2012.
- 11 Rodova M, Fu J, Watkins DN, Srivastava RK and Shankar S: Sonic hedgehog signaling inhibition provides opportunities for targeted therapy by sulforaphane in regulating pancreatic cancer stem cell self-renewal. *Plos One* 7(9): e46083, 2012.
- 12 Mueller MT, Hermann PC, Witthauer J, Rubio-Viqueira B, Leicht SF, Huber S, Ellwart JW, Mustafa M, Bartenstein P, D'Haese JG, Schoenberg MH, Berger F, Jauch KW, Hidalgo M and Heeschen C: Combined targeted treatment to eliminate tumorigenic cancer stem cells in human pancreatic cancer. *Gastroenterology* 137(3): 1102-1113, 2009.
- 13 Seidel K, Ahn CP, Lyons D, Nee A, Ting K, Brownell L, Cao T, Carano RA, Curran T, Schober M, Fuchs E, Joyner A, Martin GR, de Sauvage FJ and Klein OD: Hedgehog signaling regulates the generation of ameloblast progenitors in the continuously growing mouse incisor. *Development* 137(22): 3753-3761, 2010.
- 14 Katoh Y and Katoh M: Hedgehog target genes: mechanisms of carcinogenesis induced by aberrant hedgehog signaling activation. *Curr Mol Med* 9(7): 873-886, 2009.
- 15 Zhang C, Li C, He F, Cai Y and Yang H: Identification of CD44⁺CD24⁺ gastric cancer stem cells. *J Cancer Res Clin Oncol* 137(11): 1679-1686, 2011.
- 16 Chen JK, Taipale J, Cooper MK and Beachy PA: Inhibition of Hedgehog signaling by direct binding of cyclopamine to Smoothened. *Genes Dev* 16(21): 2743-2748, 2002.
- 17 Sims-Mourtada J, Izzo JG, Apisarnthanarax S, Wu TT, Malhotra U, Luthra R, Liao Z, Komaki R, van der Kogel A, Ajani J and Chao KS: Hedgehog: an attribute to tumor regrowth after chemoradiotherapy and a target to improve radiation response. *Clin Cancer Res* 12(21): 6565-6572, 2006.
- 18 Xu X, Zhou Y, Xie C, Wei SM, Gan H, He S, Wang F, Xu L, Lu J, Dai W, He L, Chen P, Wang X and Guo C: Genome-wide screening reveals an EMT molecular network mediated by Sonic hedgehog-Gli1 signaling in pancreatic cancer cells. *PLoS One* 7(8): e43119, 2012.
- 19 Yang J and Weinberg RA: Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. *Dev Cell* 14(6): 818-829, 2008.
- 20 Shida T, Furuya M, Nikaido T, Hasegawa M, Koda K, Oda K, Miyazaki M, Kishimoto T, Nakatani Y and Ishikura H: Sonic Hedgehog-Gli1 signaling pathway might become an effective therapeutic target in gastrointestinal neuroendocrine carcinomas. *Cancer Biol Ther* 5(11): 1530-1538, 2006.
- 21 Herschkowitz JI, Zhao W, Zhang M, Usary J, Murrow G, Edwards D, Knezevic J, Greene SB, Darr D, Troester MA, Hilsenbeck SG, Medina D, Perou CM and Rosen JM: Comparative oncogenomics identifies breast tumors enriched in functional tumor-initiating cells. *Proc Natl Acad Sci USA* 109(8): 2778-2783, 2012.
- 22 Taube JH, Herschkowitz JI, Komurov K, Zhou AY, Gupta S, Yang J, Hartwell K, Onder TT, Gupta PB, Evans KW, Hollier BG, Ram PT, Lander ES, Rosen JM, Weinberg RA and Mani SA: Core epithelial-to-mesenchymal transition interactome gene-expression signature is associated with claudin-low and metaplastic breast cancer subtypes. *Proc Natl Acad Sci USA* 107(35): 15449-15454, 2010.
- 23 Turksen K: Claudins and cancer stem cells. *Stem Cell Rev* 7(4): 797-798, 2011.
- 24 Santin AD, Cane S, Bellone S, Palmieri M, Siegel ER, Thomas M, Roman JJ, Burnett A, Cannon MJ and Pecorelli S: Treatment of chemotherapy-resistant human ovarian cancer xenografts in C.B-17/CID mice by intraperitoneal administration of Clostridium perfringens enterotoxin. *Cancer Res* 65(10): 4334-4342, 2005.

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