SAKAGUCHI AND CHIDA

Roles of Prion Protein in Virus Infections

Suehiro Sakaguchi and Junji Chida

Division of Molecular Neurobiology, Institute for Enzyme Research (KOSOKEN),

Tokushima University, Tokushima, Japan.

Running title: Prion protein in virus infections

Keywords: prion protein, oxidative stress, influenza A virus, superoxide dismutase, copper, reactive oxygen species

Address correspondence to:

Suehiro Sakaguchi, MD/PhD

Division of Molecular Neurobiology

Institute for Enzyme Research (KOSOKEN)

Tokushima University

3-18-15 Kuramoto

Tokushima 770-8503

Japan

E-mail: sakaguchi@tokushima-u.ac.jp

Abstract

The normal cellular prion protein, designated PrP^C, is a membrane glycoprotein expressed most abundantly in brains, particularly by neurons, and to a lesser extent in non-neuronal tissues including lungs. Conformational conversion of PrP^C into the amyloidogenic isoform is a key pathogenic event in prion diseases. We recently found that PrP^C has a protective role against infection with influenza A viruses (IAVs) in mice by reducing reactive oxygen species in the lungs after infection with IAVs. The anti-oxidative activity of PrP^C is probably attributable to its function to activate anti-oxidative enzyme Cu/Zn-superoxide dismutase, or SOD1, through regulating Cu content in lungs infected with IAVs. Oxidative stress could play a pivotal role in the pathogenesis of a wide range of viral infections. Here, we introduce our and others' studies on the role of PrP^C in viral infections, and raise the attractive possibility that PrP^C might be a novel target molecule for development of anti-oxidative therapeutics against not only IAV infection but also other viral infections.

Introduction

Viral infection frequently causes oxidative stress by inducing overproduction of reactive species such as reactive oxygen species (ROS) through enzymatic and non-enzymatic mechanisms in host cells (Camini et al., 2017; Li et al., 2017; Peterhans, 1997a; Schwarz, 1996). ROS are chemically reactive molecules containing oxygen, including superoxide, hydrogen peroxide, and hydroxyl radical. The overproduced ROS overly oxidizes proteins, lipids, and DNA, thereby damaging these molecules in host cells eventually contributing to the pathogenesis of virus infection. Cells are also equipped with anti-oxidative mechanisms to balance cellular redox homeostasis. Superoxide dismutase (SOD), catalase, and glutathione peroxidase are major anti-oxidative enzymes (Sgarbanti et al., 2014). Another reactive species, nitric oxide (NO), also contributes to the pathogenesis of virus infections (Perrone et al., 2013; Peterhans, 1997b). NO is produced by NO synthases (NOSs) and converted into the potent oxidative agent nitroperoxide through interaction with oxygen radicals, particularly superoxide (Akaike and Maeda, 2000). Mitigation of oxidative stress in host cells through either interfering with the oxidative mechanisms or enhancing the anti-oxidative mechanisms, or both, can be therapeutically beneficial for viral infections.

The normal cellular prion protein, designated PrP^C, is a membrane glycoprotein tethered to the outer cell membrane via a glycosylphosphatidylinositol anchor moiety and expressed most abundantly in brains, particularly by neurons, and to a lesser extent in non-neuronal tissues including hearts, kidneys, and lungs (Oesch *et al.*, 1985; Prusiner, 1998). Conformational conversion of PrP^C into the amyloidogenic isoform is a key pathogenic event

in prion diseases, a group of neurodegenerative disorders, which include Creutzfeldt-Jakob disease in humans and scrapie and bovine spongiform encephalopathy in animals (Prusiner, 1998). Several lines of evidence have suggested that PrP^C might have an anti-oxidative function. PrP^C binds to copper (Cu) ions via the histidine residues within the N-terminally located octapeptide repeat (OR) region, which is comprised of 5 tandem repeats of 8 amino acids (Jackson *et al.*, 2001). PrP^C is suggested to regulate anti-oxidative enzymes, such as Cu/Zn-SOD, or SOD1, via transfer of the bound Cu ions to the enzymes (Haigh and Brown, 2006). Other cellular functions, including cell trafficking, cell adhesion, cell differentiation, cell signaling, and cell survival, have been also suggested for PrP^C (Aguzzi *et al.*, 2008).

We recently found that PrP^C has a protective role against infection with influenza A viruses (IAVs) in mice probably through its anti-oxidative function (Chida *et al.*, 2018). There have been also several reports that PrP^C might be involved in protection against different virus infections through different mechanisms (Alais *et al.*, 2012; Baj *et al.*, 2005; Caruso *et al.*, 2009; Nakamura *et al.*, 2003; Nasu-Nishimura *et al.*, 2008; Thackray and Bujdoso, 2002). Here, we introduce our and others' studies on the role of PrP^C in virus infections.

Anti-oxidative treatments against IAV infection in mice

IAV is an enveloped, negative sense, single-stranded RNA virus, causing seasonal epidemics of influenza (Fiore *et al.*, 2008). High morbidity and mortality are observed in infected people, particularly in the young and elderly and those with underlying chronic diseases in lung or

cardiovascular systems (Fiore et al., 2008). Several lines of evidence indicate that ROS plays a pivotal role in the pathogenesis of IAV infection (Akaike et al., 1990; Oda et al., 1989; Tantcheva et al., 2003; Vlahos and Selemidis, 2014). Mice deficient in NOX2, a subunit of the ROS-producing multi-protein complex enzyme nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, showed reduced lung injuries after infection with IAV/X-31 (H3N2) and IAV/Puerto Rico/8/34 (H1N1) (hereafter referred to as IAV/PR8) (Vlahos et al., 2011). The inhibitor of another ROS-producing enzyme xanthine oxidase (XO), allopurinol, also reduced mortality of mice infected with IAV/Kumamoto/Y5/67(H2N2) (referred to as IAV/Kumamoto) (Akaike et al., 1990). These results suggest that NADPH oxidase and XO are major ROS-producing enzymes in lungs infected with IAVs, and that reducing the oxidative mechanism could be effective in treatment for IAV infection. On the contrary, it was shown that administration of pyran polymer-conjugated SOD1 successfully reduced the mortality of mice infected with IAV/Kumamoto (Oda et al., 1989). It is thus suggested that SOD1 might be a major anti-oxidative enzyme in IAV infection and that enhancing the anti-oxidative mechanisms could be also therapeutically effective against IAV infection. Treatment with the NOS inhibitor, N^{ω} -monomethyl-L-arginine, successfully reduced the mortality of mice infected with IAV/Kumamoto (Akaike et al., 1996), suggesting that NO could also play an important role in the pathogenesis of IAV infection.

Anti-oxidative role of PrPC in protection against IAV infection

We showed that PrP^C was expressed by alveolar type 1 and 2 epithelial cells (AT1 and AT2 cells) and bronchiolar Clara epithelial cells in mouse lungs (Chida *et al.*, 2018), and that mice devoid of PrP^C (*Prnp*^{0/0}) were highly susceptible to intranasal infection with IAV/PR8, A/Aichi/2/68 (H3N2), and A/WSN/33 (H1N1), with markedly elevated mortality, compared to control wild-type (WT) mice (Chida *et al.*, 2018). Infected *Prnp*^{0/0} lungs were severely damaged, with higher infiltration of inflammatory cells, higher levels of inflammatory cytokines and slightly but significantly higher virus titers than control WT lungs (Chida *et al.*, 2018). AT2 and Clara cells succumbed to apoptosis in infected *Prnp*^{0/0} lungs more than in control WT lungs (Chida *et al.*, 2018). In contrast, AT1 cells were not damaged in infected *Prnp*^{0/0} and WT lungs (Chida *et al.*, 2018). This is consistent with IAV/PR8 infection not damaging AT1 cells in C57BL/6 mice (Yamada *et al.*, 2012). These results indicate that PrP^C could have a protective role against lethal infection with IAVs in mice by mitigating lung injuries induced by IAV infection (Fig. 1).

ROS levels were higher in IAV-infected *Prnp*^{0/0} lungs compared to control WT lungs (Chida *et al.*, 2018). In addition, treatment with butylated hydroxyanisole, a ROS scavenger, decreased the mortality of infected *Prnp*^{0/0} mice to that of control WT mice (Chida *et al.*, 2018). These results suggest that PrP^C could play an anti-oxidative role to reduce ROS levels in IAV-infected lungs, thereby providing a protection against lethal infection with IAVs (Fig. 1). In contrast to higher ROS levels in infected *Prnp*^{0/0} lungs, Cu content and SOD1 activity were lower in infected *Prnp*^{0/0} lungs than in control WT mice (Chida *et al.*, 2018). It is thus conceivable that PrP^C might exert its anti-oxidative role through regulation of

the Cu content and SOD1 activity in IAV-infected lungs (Fig. 1). Tg(PrPΔOR)/*Prnp*^{0/0} mice, which express transgenic mouse PrP with a deletion of the Cu-binding OR region on the *Prnp*^{0/0} background (Yoshikawa *et al.*, 2008), also showed lower Cu content, lower SOD1 activity, and higher ROS levels in their lungs and higher mortality after infection with IAV/PR8 (Chida *et al.*, 2018). These results suggest that the Cu-binding OR region plays an important role for PrP^C to regulate the Cu content and SOD1 activity and then to exert the anti-oxidative effect in IAV-infected lungs (Fig. 1).

IAVs primarily infect lung epithelial cells, including AT2 and Clara cells, and then cause oxidative stress in them (Liu *et al.*, 2017; Sgarbanti *et al.*, 2014; Short *et al.*, 2014). *Prnp*^{0/0} epithelial cells do not sufficiently combat the oxidative stress due to lack of PrP^C, therefore undergoing apoptosis more easily than WT epithelial cells after IAV infection. The higher apoptosis of *Prnp*^{0/0} epithelial cells then provokes higher inflammatory responses leading to higher production of inflammatory cytokines in infected *Prnp*^{0/0} lungs, eventually causing higher mortality of *Prnp*^{0/0} mice after infection with IAVs.

Roles of PrP^C in other virus infections

Other groups have also investigated the roles of PrP^C in other virus infections (Alais *et al.*, 2012; Baj *et al.*, 2005; Caruso *et al.*, 2009; Nakamura *et al.*, 2003; Nasu-Nishimura *et al.*, 2008; Thackray and Bujdoso, 2002). Higher neuronal apoptosis was reported in the brains of *Prnp*^{0/0} mice than in control WT mice after infection with encephalomyocarditis virus B variant (EMCV-B), with less infiltration of inflammatory cells including microglia in infected

Prnp^{0/0} brains than in control WT brains (Nasu-Nishimura et al., 2008). EMCV-B was similarly replicated in the brains of infected Prnp^{0/0} and WT mice (Nasu-Nishimura et al., 2008). These results suggest that PrP^C might be involved in protection of neurons from EMCV-B infection-induced apoptosis possibly through activation of brain inflammatory responses against EMCV-B infection without affecting EMCV-B replication. It was also reported that PrP^C might be involved in protection against latent infection with herpes simplex virus type 1 (HSV-1) (Thackray and Bujdoso, 2002). Mice overexpressing transgenic PrP^C were sensitive to acute infection of HSV-1 strain SC16 in the central and peripheral neuronal tissues, exhibiting higher mortality than control mice (Thackray and Bujdoso, 2002). However, latent infection of the virus in these tissues was significantly suppressed in these mice (Thackray and Bujdoso, 2002). Lower induction of autophagy was reported in *Prnp*^{0/0} astrocytes than in WT astrocytes after infection with HSV-1 strain 17, suggesting that PrP^C might be involved in induction of autophagy in astrocytes after infection with HSV-1 (Korom et al., 2013). However, it remains to be determined whether or not the enhanced acute infection of HSV-1 and the suppressed latent infection of HSV-1 in PrP^C-overexpressing mice can be attributable to the higher induction of autophagy in HSV-1-infected astrocytes. It was also reported that PrP^C inhibited production of human immunodeficiency virus type 1 (HIV-1) in cultured cells transfected with an infectious HIV-1 molecular clone (Alais et al., 2012). PrP^C disturbed translation of the HIV-1 genomic RNA probably through binding to the genomic RNA (Gabus et al., 2001). It has been further reported that PrP^C might be involved in protection against infection with coxsackievirus B3 (Nakamura *et al.*, 2003), adenovirus 5 (Caruso *et al.*, 2009), and poliovirus-1 (Baj *et al.*, 2005).

Perspectives

We showed that PrP^C has a protective role against lethal infection with IAVs in mice by exerting ant-oxidative activity (Chida et al., 2018). Worldwide spread of IAVs, which are resistant to the currently available anti-influenza agents, has raised great health concerns about pandemics with these resistant IAVs among human populations (Hurt et al., 2009; McKimm-Breschkin et al., 1998; Mishin et al., 2005). The currently available agents such as neuraminidase inhibitors target the molecules encoded by IAVs, promoting the emergence of the IAVs carrying mutations in the genes encoding the targeted molecules and eventually propagating these resistant mutant IAVs among human populations (Hurt et al., 2009; McKimm-Breschkin et al., 1998; Mishin et al., 2005). Therefore, host molecules involved in protection against IAV infection would be plausible targets for development of anti-influenza agents because the agents targeting host molecules are considered not to induce resistant IAVs. Anti-oxidative therapeutics against IAV infection, by targeting the ROS-generating enzymes or by administrating anti-oxidants or anti-oxidant enzymes, has been shown to successfully protect mice from lethal infection with IAVs (Akaike et al., 1990; Oda et al., 1989; Tantcheva et al., 2003; Vlahos and Selemidis, 2014). Our current findings suggest that PrP^C is a new target molecule for anti-oxidative therapeutics against IAV infection. It has been reported that PrP^C protected neurons from anisomycin-induced apoptosis via interaction

with stress-inducible protein 1 (STI1), a STI1-derived peptide, or anti-PrP antibodies (Chiarini *et al.*, 2002; Zanata *et al.*, 2002), and that the interaction with STI1 could be involved in PrP^C-dependent activation of SOD (Sakudo *et al.*, 2005). It is thus interesting to investigate whether these ligands could elicit the anti-oxidative activity of PrP^C and protect against IAV infection and other virus infections, in which oxidative stress plays a pivotal role in the pathogenesis.

Acknowledgments

This study was partly supported to SS by JSPS KAKENHI Grant Number 26293212 and 15K15380, and Grant-in-Aid for Scientific Research on Innovative Areas (Brain Protein Aging and Dementia Control) Grant Number 15H01560 and 17H05701 from MEXT, and to JC by JSPS KAKENHI Grant Number 16K10029.

Disclosure Statement

No competing financial interests exist.

References

Aguzzi, A., Baumann, F., Bremer, J. (2008). The prion's elusive reason for being. Annu Rev Neurosci **31**, 439-477.

10

- Akaike, T., Ando, M., Oda, T., Doi, T., Ijiri, S., Araki, S., *et al.* (1990). Dependence on O2-generation by xanthine oxidase of pathogenesis of influenza virus infection in mice. J Clin Invest **85**, 739-745.
- Akaike, T., Maeda, H. (2000). Nitric oxide and virus infection. Immunology 101, 300-308.
- Akaike, T., Noguchi, Y., Ijiri, S., Setoguchi, K., Suga, M., Zheng, Y.M., *et al.* (1996).

 Pathogenesis of influenza virus-induced pneumonia: involvement of both nitric oxide and oxygen radicals. Proc Natl Acad Sci USA **93**, 2448-2453.
- Alais, S., Soto-Rifo, R., Balter, V., Gruffat, H., Manet, E., Schaeffer, L., *et al.* (2012). Functional mechanisms of the cellular prion protein (PrP(C)) associated anti-HIV-1 properties. Cell Mol Life Sci **69**, 1331-1352.
- Baj, A., Bettaccini, A., Nishimura, T., Onodera, T., Toniolo, A. (2005). Poliovirus type 1 infection of murine PRNP-knockout neuronal cells. J Neurovirol **11**, 237-246.
- Camini, F.C., da Silva Caetano, C.C., Almeida, L.T., de Brito Magalhaes, C.L. (2017).

 Implications of oxidative stress on viral pathogenesis. Arch Virol **162**, 907-917.
- Caruso, P., Burla, R., Piersanti, S., Cherubini, G., Remoli, C., Martina, Y., et al. (2009). Prion expression is activated by Adenovirus 5 infection and affects the adenoviral cycle in human cells. Virology **385**, 343-350.
- Chiarini, L.B., Freitas, A.R., Zanata, S.M., Brentani, R.R., Martins, V.R., Linden, R. (2002).

 Cellular prion protein transduces neuroprotective signals. Embo J **21**, 3317-3326.

- Chida, J., Hara, H., Yano, M., Uchiyama, K., Das, N.R., Takahashi, E., et al. (2018). Prion protein protects mice from lethal infection with influenza A viruses. PLoS Pathog 14, e1007049.
- Fiore, A.E., Shay, D.K., Broder, K., Iskander, J.K., Uyeki, T.M., Mootrey, G., *et al.* (2008).

 Prevention and control of influenza: recommendations of the Advisory Committee on Immunization Practices (ACIP), 2008. MMWR Recommendations and reports:

 Morbidity and mortality weekly report Recommendations and reports/Centers for Disease Control **57**, 1-60.
- Gabus, C., Derrington, E., Leblanc, P., Chnaiderman, J., Dormont, D., Swietnicki, W., et al. (2001). The prion protein has RNA binding and chaperoning properties characteristic of nucleocapsid protein NCP7 of HIV-1. J Biol Chem 276, 19301-19309.
- Haigh, C.L., Brown, D.R. (2006). Prion protein reduces both oxidative and non-oxidative copper toxicity. J Neurochem **98**, 677-689.
- Hurt, A.C., Holien, J.K., Parker, M., Kelso, A., Barr, I.G. (2009). Zanamivir-resistant influenza viruses with a novel neuraminidase mutation. J Virol **83**, 10366-10373.
- Jackson, G.S., Murray, I., Hosszu, L.L., Gibbs, N., Waltho, J.P., Clarke, A.R., et al. (2001).
 Location and properties of metal-binding sites on the human prion protein. Proc Natl
 Acad Sci USA 98, 8531-8535.
- Korom, M., Wylie, K.M., Wang, H., Davis, K.L., Sangabathula, M.S., Delassus, G.S., *et al.* (2013). A proautophagic antiviral role for the cellular prion protein identified by infection with a herpes simplex virus 1 ICP34.5 mutant. J Virol **87**, 5882-5894.

- Li, Z., Xu, X., Leng, X., He, M., Wang, J., Cheng, S., *et al.* (2017). Roles of reactive oxygen species in cell signaling pathways and immune responses to viral infections. Arch Virol **162**, 603-610.
- Liu, M., Chen, F., Liu, T., Chen, F., Liu, S., Yang, J. (2017). The role of oxidative stress in influenza virus infection. Microbes Infect 19, 580-586.
- McKimm-Breschkin, J.L., Sahasrabudhe, A., Blick, T.J., McDonald, M., Colman, P.M., Hart, G.J., *et al.* (1998). Mutations in a conserved residue in the influenza virus neuraminidase active site decreases sensitivity to Neu5Ac2en-derived inhibitors. J Virol **72**, 2456-2462.
- Mishin, V.P., Hayden, F.G., Gubareva, L.V. (2005). Susceptibilities of antiviral-resistant influenza viruses to novel neuraminidase inhibitors. Antimicrob Agents Chemother **49**, 4515-4520.
- Nakamura, Y., Sakudo, A., Saeki, K., Kaneko, T., Matsumoto, Y., Toniolo, A., et al. (2003).

 Transfection of prion protein gene suppresses coxsackievirus B3 replication in prion protein gene-deficient cells. J Gen Virol 84, 3495-3502.
- Nasu-Nishimura, Y., Taniuchi, Y., Nishimura, T., Sakudo, A., Nakajima, K., Ano, Y., *et al.* (2008). Cellular prion protein prevents brain damage after encephalomyocarditis virus infection in mice. Arch Virol **153**, 1007-1012.
- Oda, T., Akaike, T., Hamamoto, T., Suzuki, F., Hirano, T., Maeda, H. (1989). Oxygen radicals in influenza-induced pathogenesis and treatment with pyran polymer-conjugated SOD. Science **244**, 974-976.

- Oesch, B., Westaway, D., Walchli, M., McKinley, M.P., Kent, S.B., Aebersold, R., *et al.* (1985). A cellular gene encodes scrapie PrP 27-30 protein. Cell **40**, 735-746.
- Perrone, L.A., Belser, J.A., Wadford, D.A., Katz, J.M., Tumpey, T.M. (2013). Inducible nitric oxide contributes to viral pathogenesis following highly pathogenic influenza virus infection in mice. J Infect Dis **207**, 1576-1584.
- Peterhans, E. (1997a). Oxidants and antioxidants in viral diseases: disease mechanisms and metabolic regulation. J Nutr **127**, 962S-965S.
- Peterhans, E. (1997b). Reactive oxygen species and nitric oxide in viral diseases. Biol Trace Elem Res **56**, 107-116.
- Prusiner, S.B. (1998). Prions. Proc Natl Acad Sci USA 95, 13363-13383.
- Sakudo, A., Lee, D.C., Li, S., Nakamura, T., Matsumoto, Y., Saeki, K., *et al.* (2005). PrP cooperates with STI1 to regulate SOD activity in PrP-deficient neuronal cell line. Biochem Biophys Res Commun **328**, 14-19.
- Schwarz, K.B. (1996). Oxidative stress during viral infection: a review. Free Radic Biol Med **21**, 641-649.
- Sgarbanti, R., Amatore, D., Celestino, I., Marcocci, M.E., Fraternale, A., Ciriolo, M.R., *et al.* (2014). Intracellular redox state as target for anti-influenza therapy: are antioxidants always effective? Curr Top Med Chem **14**, 2529-2541.
- Short, K.R., Kroeze, E.J., Fouchier, R.A., Kuiken, T. (2014). Pathogenesis of influenza-induced acute respiratory distress syndrome. Lancet Infect Dis 14, 57-69.

- Tantcheva, L.P., Stoeva, E.S., Galabov, A.S., Braykova, A.A., Savov, V.M., Mileva, M.M. (2003). Effect of vitamin E and vitamin C combination on experimental influenza virus infection. Methods Find Exp Clin Pharmacol **25**, 259-264.
- Thackray, A.M., Bujdoso, R. (2002). PrP(c) expression influences the establishment of herpes simplex virus type 1 latency. J Virol **76**, 2498-2509.
- Vlahos, R., Selemidis, S. (2014). NADPH oxidases as novel pharmacologic targets against influenza A virus infection. Mol Pharmacol **86**, 747-759.
- Vlahos, R., Stambas, J., Bozinovski, S., Broughton, B.R., Drummond, G.R., Selemidis, S. (2011). Inhibition of Nox2 oxidase activity ameliorates influenza A virus-induced lung inflammation. PLoS Pathog 7, e1001271.
- Yamada, Y., Limmon, G.V., Zheng, D., Li, N., Li, L., Yin, L., et al. (2012). Major shifts in the spatio-temporal distribution of lung antioxidant enzymes during influenza pneumonia. PLoS ONE 7, e31494.
- Yoshikawa, D., Yamaguchi, N., Ishibashi, D., Yamanaka, H., Okimura, N., Yamaguchi, Y., et al. (2008). Dominant-negative effects of the N-terminal half of prion protein on neurotoxicity of prion protein-like protein/doppel in mice. J Biol Chem 283, 24202-24211.
- Zanata, S.M., Lopes, M.H., Mercadante, A.F., Hajj, G.N., Chiarini, L.B., Nomizo, R., *et al.* (2002). Stress-inducible protein 1 is a cell surface ligand for cellular prion that triggers neuroprotection. Embo J **21**, 3307-3316.

Figure legends

1

- 2 FIG. 1. A possible mechanism for the protective role of PrP^C against IAV infection-induced
- 3 apoptosis. IAV infection in lung epithelial cells causes overproduction of ROS by directly or
- 4 indirectly inducing production of inflammatory cytokines, leading to apoptosis in infected
- 5 epithelial cells. PrP^C could regulate the enzymatic activity of SOD1 by transferring Cu ions,
- 6 which are bound to the N-terminal OR region, thereby mitigating the burden of ROS in
- 7 infected cells and eventually protecting the cells from undergoing apoptosis.

