

This version of the article has been accepted for publication, after peer review (when applicable) and is subject to Springer Nature's AM terms of use (<https://www.springernature.com/gp/open-research/policies/accepted-manuscript-terms>), but is not the Version of Record and does not reflect post-acceptance improvements, or any corrections. The Version of Record is available online at: <https://doi.org/10.1007/s12185-020-02975-x>

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Proposed type of manuscript

Original article

Title

Involvement of oral bacteria and oral immunity as risk factors for chemotherapy-induced fever with neutropenia in patients with hematological cancer

Yuka Sogawa^{1, 2}, Makoto Fukui¹, Shingen Nakamura³, Kimiko Sogabe³, Ryohei Sumitani³, Masami Yoshioka², Masahiro Abe³, Daisuke Hinode^{1*}

¹ Department of Hygiene and Oral Health Science, Tokushima University Graduate School of Biomedical Sciences, Tokushima 770-8504, Japan

² Tokushima Bunri University Faculty of Health and Welfare, Nishihama, Yamashiro-cho, Tokushima, 770-8514 Japan

³ Department of Hematology, Endocrinology, and Metabolism, Tokushima University Graduate School of Biomedical Sciences, Tokushima 770-8503, Japan

*Corresponding author

Dr. Daisuke Hinode

Department of Hygiene and Oral Health Science, Tokushima University Graduate School of Biomedical Sciences, Tokushima 770-8504, Japan

Telephone: +81-88-633-7543

Fax: +81-88-633-7543

E-mail: hinode@tokushima-u.ac.jp

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

The running title

Risk factors for chemotherapy-induced fever with neutropenia

1
2
3 **Abstract**
4

5 The aim of this study is to investigate the association between chemotherapy-induced
6 **fever with neutropenia less than 1,500/ μ L (FwN)** and oral bacteria and/or oral immunity
7
8 in patients with hematological cancer. Thirty-two patients with hematological cancer
9
10 were enrolled in the study. Secretory immunoglobulin A (sIgA) in saliva and the
11
12 anaerobic bacteria in tongue coating of each subject was assessed before the first
13
14 chemotherapy. Eleven subjects had an onset of **FwN** and 21 subjects did not during the
15
16 observation periods. It was revealed by the cox-proportional hazard model analysis that
17
18 the levels of sIgA were low (HR 0.98, $p < 0.05$) and the rate of *Fusobacterium nucleatum*
19
20 [*F. nucleatum* count per total bacterial count (%)] was high (HR 1.65, $p < 0.05$) in
21
22 patients with **FwN** onset. Using ROC curve analysis, the optimal cut-off point based on
23
24 the AUC in the *F. nucleatum* / sIgA ratio was 0.023, and this model had a 78.4%
25
26 probability ($p < 0.01$). The risk of **FwN** onset was also significantly higher among the
27
28 group of ≥ 0.023 *F. nucleatum* / sIgA ratio (HR 66.06, $p < 0.01$). These results suggest
29
30 that the rate of *F. nucleatum* and the levels of sIgA at baseline might be related to **FwN**
31
32 onset as risk factors.
33
34
35
36
37
38
39
40
41
42
43
44

45 **Keyword**
46

47 **Fever with neutropenia**, Hematological cancer, Risk factor, Secretory immunoglobulin
48
49 A, *Fusobacterium nucleatum*
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 **Introduction**
4

5 It has been reported that adverse events observed in cancer chemotherapy were
6 caused by a combination of many complex factors. Among them, febrile neutropenia
7 (FN) is one of the frequent adverse events of hematological cancer chemotherapy.
8 Neutrophils, a type of white blood cell, take in pathogens that invade the body, digest
9 and degrade them with enzymes released from granules in the cytoplasm. When
10 chemotherapy is performed on patients with hematological cancer, bone marrow
11 function decreases, and neutropenia is observed about seven days after drug
12 administration. Notably, there is a high risk of fever due to susceptibility to infection
13 during neutropenia [1].
14
15
16
17
18
19
20
21
22
23
24
25

26 FN is defined as a condition in which the axillary temperature is 37.5°C (or oral
27 temperature 38°C) or higher. Also, the number of neutrophil count is less than 500
28 cells/ μ L or is less than 1,000/ μ L with a predicted decline to 500/ μ L within 48 hours [2].
29 In particular, it has been reported that cancer chemotherapy with strong bone marrow
30 suppression, such as in the treatment of hematological cancers, increases the likelihood
31 of FN becoming severe, leading to death [3]. On the other hand, there is a risk that local
32 infections due to oral mucositis may progress to serious infections [4].
33 Granulocytopenia, which occurs at the same time as oral mucositis, increases the risk of
34 bacteremia and sepsis. Gram-negative bacilli, the normal bacterial flora in the mouth,
35 can spread hematogenously from the site of the ulcer and can cause local or systemic
36 infection. Anaerobic bacteria represented by periodontopathogenic bacteria existing in
37 the oral cavity are also assumed to be the infection source in those cases.
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54

55 There were few studies regarding the association between FN development and the
56 oral cavity. It has been reported that professional oral care significantly reduced the
57
58
59
60
61
62
63
64
65

1
2
3 incidence of FN [5] and that *Fusobacterium nucleatum*, a periodontopathogenic bacteria,
4
5 was the cause of septic patients with FN [6]. However, the details of the relationship
6
7 between the onset of FN and oral bacteria are still unknown, and it has not yet been
8
9 clarified in many ways as to what kinds of microorganisms are actively involved in its
10
11 progress. Professional oral care management has been widely performed to prevent
12
13 adverse oral events in patients with hematological cancers on intensive chemotherapies.
14
15 However, the therapeutic efficacy of professional oral care approaches to prevent the
16
17 incidence of FN remains elusive.
18
19
20

21
22 On the other hand, it has been demonstrated that secretory IgA (sIgA) plays an
23
24 important role as the “first line of defense” of the oral mucosal surface as an
25
26 antibacterial factor in saliva. It was reported that the amount of sIgA and saliva volume
27
28 was reduced by chemotherapy for patients with breast cancer [7]. Furthermore,
29
30 Karolewska et al. reported that the amount of sIgA decreased during chemotherapy for
31
32 children with acute leukemia, and sIgA levels in patients with oral mucositis were lower
33
34 than that in patients without oral mucositis [8]. Thus, cancer treatment also affects oral
35
36 immunity.
37
38
39

40
41 The purpose of this study was to 1) investigate the incidence of fever with
42
43 neutropenia (FwN) among adverse events in hematological cancer patients receiving
44
45 chemotherapy, 2) to analyze the association with FwN and oral bacteria such as
46
47 anaerobic microorganisms and other factors in the oral cavity, especially, sIgA, and 3) to
48
49 propose predictive indices highly relevant to FwN onset.
50
51
52
53
54

55 **Material and Methods**

56 **1. Subjects**

1
2
3 Thirty-three patients aged 22 to 86 years old (20 males and 13 females) whose oral
4
5 cavity status were checked and who received their first chemotherapy for hematological
6
7 cancer at Tokushima University Hospital from April 2012 to December 2016 were
8
9 initially enrolled in the study. Before enrollment, the participants were informed about
10
11 the methods and objectives of the study, and they provided written informed consent.
12
13
14
15
16

17 **2. Clinical parameters**

18
19 Diagnosis and treatment procedure of hematological cancer, the chemotherapeutic
20
21 drug used, administration of antibiotics and/or G-CSF for the prevention or treatment of
22
23 infections, the systemic conditions including diabetes mellitus were obtained from the
24
25 patient's medical record. The period of time including the presence of fever, and results
26
27 of the blood test, such as the value of serum albumin and neutrophil count (or the white
28
29 blood cell count), were also obtained. **In this study, FwN was set as a fever of unknown**
30
31 **origin (an axillary temperature of 37.5°C or higher) with neutropenia less than 1,500/ μ L**
32
33 **to widely analyze the patients enrolled.** FN is defined as a fever of unknown origin with
34
35 neutropenia, a neutrophil count of less than 500/ μ L, and an axillary temperature of
36
37 37.5°C or higher based on the Japanese Society of Medical Oncology guidelines [9].
38
39 The day of the onset of **FwN** was also confirmed from the patient's medical record.
40
41 Observation periods were decided 28 days from the first day of the chemotherapy. Also,
42
43 the smoking habits of the patients were recorded at baseline.
44
45
46
47
48
49
50
51
52

53 **3. Assessment of samples of the saliva and the tongue coating**

54
55 Saliva and tongue coating samples from each subject were assessed before the first
56
57 chemotherapy. Unstimulated whole saliva from each subject was collected using a
58
59
60
61
62
63
64
65

1
2
3 sterilized cotton swab kit (Salikids®; Saersted, Vümbrecht, Germany) before lunch. The
4
5 cotton roll was placed into the oral cavity of each subject for 3 min, then the roll was
6
7 returned into the Salikids® tube, and was centrifuged at 3,000 x g for 2 min at 4°C. The
8
9 resulting aliquots were divided into analytical tubes and were kept in a deep freezer at
10
11 -80°C until use.
12
13

14
15 The levels of salivary protein were measured using Protein Assay Kit (Bio-Rad,
16
17 Hercules, CA, USA) according to the method of Bradford [10] with BSA as a standard.
18
19 The levels of sIgA of saliva were measured using an enzyme immune-assay kit (EIA
20
21 s-IgA test, Medical & Biological Laboratories, Nagoya, Japan) according to the
22
23 manufacturer's instructions. Both of the levels of salivary protein and sIgA were
24
25 evaluated as the total amount per minute secretion.
26
27

28
29 Bacterial Counter™ (Panasonic, Osaka, Japan) was used to assess the number of
30
31 bacteria in tongue coating according to the manufacturer's instructions. Tongue coating
32
33 samples were collected using a sterile 5mm-diameter cotton stick by swabbing the
34
35 tongue dorsum three times from back to front (approx. 2-cm-long swabbing motions).
36
37 Samples were suspended in 5 ml of distilled water in disposable cups and bacterial
38
39 quantification with Bacterial Counter™ was performed. Thereafter, the samples were
40
41 dispensed into vials and kept at -80°C until used.
42
43
44

45
46 Tongue coating samples were also used to quantify the number of three
47
48 periodontopathogenic bacteria (*Porphyromonas gingivalis*, *Fusobacterium nucleatum*,
49
50 and *Campylobacter rectus*) by quantitative PCR as previously reported by Moriyama et
51
52 al [11]. The MiniOpticon system (Bio-Rad Laboratories, Hercules, CA, USA) with
53
54 SYBR Green I dye was used for the quantitative PCR analysis. The primers used for the
55
56 quantitative PCR have been previously described [12]. A standard curve was generated
57
58
59
60

1
2
3 based on the known number of *F. nucleatum* ATCC 23726, *P. gingivalis* ATCC 33277,
4
5 and *C. rectus* ATCC 33238. Ten-fold serial dilutions of bacterial standards were
6
7 prepared and each extracted DNA was used. The concentrations of each organism in
8
9 tongue coating samples were calculated from the number of copies of the target
10
11 sequence.
12
13

14 15 16 17 **4. Statistical analysis**

18
19 Data were analyzed using the software IBM SPSS Statistics ver. 26 (SPSS Japan Inc.
20
21 Tokyo). The difference between the two groups with baseline was assessed using the
22
23 chi-square test or Fisher's exact test for categorical data, and the Mann-Whitney U test
24
25 for the other data. Covariates related to FwN onset was evaluated by the method of the
26
27 cox-proportional hazard model. The items of the values of $p < 0.05$ were used for the
28
29 receiver operating characteristic (ROC) curve analysis for FwN onset.
30
31

32
33 The ROC curve is the plot that displays the full picture of the trade-off between the
34
35 sensitivity (true positive rate) and (1- specificity) (false positive rate) across a series of
36
37 cut-off points. The area under the ROC curve (AUC) is considered an effective measure
38
39 of the inherent validity of a diagnostic test. Total AUC is a single index for measuring
40
41 the performance of a test [13]. The larger the AUC, the better is the overall performance
42
43 of the diagnostic test to correctly identify FwN subjects and non-FwN subjects.
44
45

46
47 To obtain the optimal cut-off points, the method uses the square of the distance
48
49 between the point (0, 1) on the upper left-hand corner of ROC space and any point on
50
51 ROC curve ie. $d^2 = (1 - \text{sensitivity})^2 + (1 - \text{specificity})^2$, then the square of this distance is
52
53 minimized [13]. The effectiveness of the optimal cut-off value obtained from the ROC
54
55 curve analysis was evaluated by the method of the cox-proportional hazard model.
56
57
58
59

1
2
3 Statistical significance was set at $p < 0.05$.
4
5
6

7 **5. Ethics**

8
9
10 The Ethics Committee of Tokushima University Hospital approved this study (protocol
11 approval number 1563).
12
13
14

15 16 17 **Results**

18 19 **1. Baseline characteristics of FwN subjects and non-FwN subjects**

20
21
22 Among the 33 subjects, one patient was excluded from this study because of fever
23 and insufficient samples at baseline. The primary diseases of hematological cancer of
24 the subjects in this study are shown in Table 1. By investigating the 32 subjects (mean
25 age 60.2 ± 15.4 years), 75% were diagnosed as having malignant lymphoma. All of the
26 subjects did not have severe stomatitis at baseline.
27
28
29
30
31
32

33
34 Table 2 summarizes the participant characteristics according to the status of FwN
35 onset such as demographic variables, clinical parameters at baseline, and regimen for
36 chemotherapy. It was revealed that 11 subjects had an onset of FwN (FwN subjects) and
37 21 subjects did not (non-FwN subjects) during observation periods (maximum 28 days).
38
39 It was observed that the age of the FwN subjects was significantly younger than those of
40 the non-FwN subjects. Also, a significant difference was observed in the levels of sIgA
41 between the two groups ($p < 0.05$).
42
43
44
45
46
47
48
49

50
51 Upon investigation of oral bacteria at baseline, the rate of *F. nucleatum* [*F. nucleatum*
52 count per total bacterial count (%)] was shown to have the tendency to be higher in the
53 FwN subjects than those in the non-FwN subjects ($p = 0.088$), whereas the rate of *P.*
54 *gingivalis* and *C. rectus* did not show any tendencies.
55
56
57
58
59
60
61
62
63
64
65

1
2
3 In contrast, results showed a relationship between salivary components and the
4 subsequent onset of FwN. The salivary volume per minute did not differ between the
5 two groups. The levels of sIgA were significantly lower in FwN subjects than in
6 non-FwN subjects ($p < 0.05$). No significant difference in the levels of salivary protein
7 was observed between the two groups.
8
9
10
11
12
13
14
15
16

17 **2. Factors related to FwN onset during chemotherapy**

18
19 As shown in Table 2, no relation of FwN onset was observed in the items of
20 antibiotics and G-CSF. The relationship between the presence of moderate neutropenia
21 (<1,500 / μ L) and the onset of FwN during the observation period were investigated. The
22 rate of moderate neutropenia during chemotherapy in the FwN subjects and in the
23 non-FwN subjects was 11 of 11 patients and 15 of 21 patients, respectively. No
24 significant difference was observed between the two groups (Fisher's exact test,
25 $p=0.071$). The difference in age between the subjects with moderate neutropenia and the
26 subjects without neutropenia was also investigated. There was no significant difference
27 in age between the two groups (Mann-Whitney U test, $p=0.29$).
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43

44 **3. Multivariable analysis by the cox-proportional hazard regression model** 45 **depicting FwN onset**

46
47
48 The results of the cox-proportional hazard regression analysis adjusted for the
49 covariates are shown in Table 3. Covariates of both the levels of sIgA and the rate of *F.*
50 *nucleatum* [*F. nucleatum* count per total bacterial count (%)] showed a significant effect
51 on FwN onset after adjusting for potential confounding effects of other selected factors
52 such as gender, age, smoking habit, diabetes mellitus, albumin, use of G-CFS and
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 regimens of the study. It was revealed that the levels of sIgA were low (HR 0.984,
4
5 95%CI: 0.970-0.999, p = 0.038) and the rate of *Fusobacterium nucleatum* [*F. nucleatum*
6
7 count per total bacterial count (%)] was high (HR 1.647, 95%CI: 1.071-2,008, p =
8
9 0.017) in patients with FwN onset.
10
11
12
13

14 **4. ROC curve analysis and the optimal cut-off value**

15
16
17 Regarding the ROC curve, the farther the curve is to the upper left corner of the graph,
18
19 the better the items are in predicting FwN onset. As shown in Fig. 1 and Table 4, based
20
21 on the AUC, the model using the levels of sIgA and the rate of *F. nucleatum* had a
22
23 74.5% probability and 68.8% probability of predicting FwN onset, respectively. In
24
25 addition, the model using the *F. nucleatum* / sIgA ratio had a 78.4% probability (p =
26
27 0.003). Subsequently, the optimal cut-off value was determined in concordance with the
28
29 FwN onset based on the AUC. As shown in Table 4, the optimal cut-off point based on
30
31 the AUC in the *F. nucleatum* / sIgA ratio was 0.023, which had a sensitivity and
32
33 specificity of 0.72 and 0.79, respectively.
34
35
36
37
38
39
40

41 **5. The hazard of FwN onset based on *F. nucleatum* / sIgA ratio**

42
43 The results of the cox-proportional hazard regression analysis adjusted for the
44
45 covariates are shown in Table 5. Covariates of *F. nucleatum* / sIgA ratio showed a
46
47 significant effect on FwN onset after adjusting for potential confounding effects of other
48
49 selected factors. The risk of FwN onset was significantly higher among the group of \geq
50
51 0.023 *F. nucleatum* / sIgA ratio compared with the group of <0.023 (HR 66.062. 95%
52
53 CI: 3.645 - 1197.37, p = 0.005).
54
55
56

57 The treatment of patients with acute myelogenous leukemia is often associated with
58
59

1
2
3 severe long-term neutropenia, which may cause disease bias. Therefore, we reevaluated
4
5 a cox-proportional hazards regression analysis without patients with myelogenous
6
7 leukemia. However, those with ≥ 0.023 *F. nucleatum* / sIgA ratio remained to be a
8
9 significantly high-risk population of FwN onset (Supplemental Table 1).
10

11
12 Figure 2 shows the cumulative hazard probability curves of FwN event stratified by
13
14 gender and age with the proportional hazard model using the *F. nucleatum* / sIgA ratio
15
16 (cut-off point: 0.023). Apparent higher FwN events were detected among the subject's
17
18 group of ≥ 0.023 *F. nucleatum* / sIgA ratio.
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Discussion

In this study, we focused on blood cancer patients with reduced immunity and investigated the relationship between FwN development and oral bacteria, in which some of the microorganisms are strongly involved in FN development [14]. It is reported that about 500 kinds of oral bacteria exist in the oral cavity and that there are 1.0×10^8 bacteria per 1mL of saliva [15]. Our study revealed that *F. nucleatum* might be a possible risk factor of chemotherapy-induced FwN in patients with hematological cancer. This organism is a gram-negative obligate anaerobic bacillus, and a type of periodontopathogenic bacterium resident in the human oral cavity [16]. Furthermore, *F. nucleatum* plays a central role in the formation of dental plaque and possesses the ability to co-aggregate with a large number of bacterial species [17]. In particular, it is abundant not only in subgingival plaque but also in supragingival plaque. As plaque matures, the environment in plaque changes, and *F. nucleatum* increases [18]. This organism is also frequently isolated from tongue coating regardless of periodontal condition [16, 19]. It was reported by Gibbons et al. [20] that among the distribution of oral bacteria, the proportion of *Fusobacterium* found at the surface of tongue dorsum was 1%. An average of 0.6% of *F. nucleatum* is detected in the saliva of healthy subjects [21]. The results of this study showed that the median of *F. nucleatum* (%) in the FwN subjects was 2.7%, and it was high when compared to those of non-FwN subjects (0.2%).

The reason for this phenomenon is unclear. An explanation to account for this observation is that it will be related to the characteristics of blood cancer, such as malaise, nausea, and vomiting, resulting in insufficient brushing. This might change the bacterial flora in the oral cavity including the severity of tongue coating. *F. nucleatum*

1
2
3 may possess the ability to co-aggregate and attract other anaerobic bacteria. These
4
5 bacteria possess an endotoxin which is one of the causes of fever. Also, these bacteria,
6
7 including *F. nucleatum*, induces apoptotic cell death in peripheral blood mononuclear
8
9 cells and polymorphonuclear cells as having an immunosuppressive role [16, 22]. These
10
11 abilities of *F. nucleatum* might cause FwN development.
12
13

14
15 It was demonstrated that *F. nucleatum* is linked to systemic diseases. For example,
16
17 this organism is known to be among the bacterium causing aspiration pneumonia [23].
18
19 Han et al. reported that *F. nucleatum* may induce premature and term stillbirths in an
20
21 in-vitro study and reviewed the epidemiological and mechanistic evidence of the role of
22
23 *F. nucleatum* in adverse pregnancy outcomes [24]. It was also reported to be related to
24
25 colorectal cancer [25] and esophageal cancer [26]. It was also reported to cause
26
27 bacteremia over a 10-month observation period [6].
28
29
30

31
32 It was not possible to observe the detailed status of oral mucositis during
33
34 chemotherapy in this study whereas the condition of each subject at baseline was
35
36 observed. It was reported that severe oral mucositis also affected the onset of FN [4].
37
38 Oral mucositis often appears during chemotherapy. The oral mucosa appears to be weak
39
40 even with slight oral mucositis. Most patients receiving chemotherapy have the side
41
42 effects of myelosuppression, therefore, it is considered that the oral environment easily
43
44 causes bacterial translocation, which is an invasion of bacteria due to physiological
45
46 abnormality. It has been reported that it is possible for *F. nucleatum* to invade the human
47
48 oral epithelial cells [27] and that the FadA adhesin/invasin conserved in *F. nucleatum* is
49
50 a key virulence factor [28]. Also, most patients undergoing chemotherapy have side
51
52 effects of myelosuppression. Therefore, this organism may also invade blood circulation,
53
54 cause infection, then cause fever. It was reported that these obligate anaerobic bacteria
55
56
57
58
59
60
61
62
63
64
65

1
2
3 have accounted for 3.4% of bacteremia in neutropenic patients [29]. It is considered that
4
5 anaerobic bacteria containing *F. nucleatum* can cause acute infection during
6
7 myelosuppression, and cause bacteremia easily. Both the bacterial translocation of *F.*
8
9 *nucleatum* and its invasive ability into oral epithelial cells might cause infection in the
10
11 blood circulation in patients with neutropenia due to cancer chemotherapy. However, *F.*
12
13 *nucleatum* was not detected in blood cultures in FwN patients in this study. *F. nucleatum*
14
15 is a well-known fastidious anaerobe to be hardly cultivated. Hence, this organism is
16
17 frequently missed in routine culture conditions employed by hospital laboratories [28].
18
19 the detection of this anaerobic bacterium using the real-time PCR method should be
20
21 implemented in the risk evaluation of FN onset in the future.
22
23
24
25

26
27 In our previous observational study, professional oral health care (POHC) was
28
29 effective for the prevention of severe oral mucositis in patients undergoing autologous
30
31 hematopoietic stem cell transplant with blood cancer undergoing chemotherapy [30].
32
33 Also, POHC significantly reduced the incidence of FN in patients treated with
34
35 allogeneic bone marrow transplantation [5]. Taken together, POHC given by dentists
36
37 and dental hygienists for patients with blood cancer before and after chemotherapy is
38
39 very important for the prevention of FN onset.
40
41
42

43
44 In the duration of neutropenia after chemotherapy, patients with malignant lymphoma
45
46 had a relatively high risk of developing FN, with the incidence of FN ranging from 12
47
48 to 23% [31]. The FwN onset among patients with malignant lymphoma in this study
49
50 were 20.8%, which is within the range reported above. In contrast, it was reported that
51
52 elderly people are at a higher risk for FN onset [14]; however, an unexpected result was
53
54 obtained in this study. FwN onset caused by myelosuppression was also found in young
55
56 subjects. One of the reasons may be that some of the patients with FwN had already
57
58
59
60
61
62
63
64
65

1
2
3 weakened resistance such as those with acute leukemia. Further study is needed to
4
5 clarify these results by increasing the number of subjects.
6

7
8 In this study, we also measured sIgA levels in saliva. As a result, the levels of sIgA
9
10 was significantly lower in the FwN subjects. The reason why the levels of sIgA were
11
12 lower in the FwN group in this study is unknown. Sun et al. have reported that patients
13
14 suffering from malignant tumors had a lower level of sIgA than healthy subjects. Also,
15
16 the level of sIgA in the hematopoietic system tumor was significantly lower than that in
17
18 other malignant tumors [32]. It is presumed that the sIgA level before cancer
19
20 chemotherapy was related to the subsequent risk of FwN onset.
21
22

23
24 Immunologic defense in the mouth is mediated by a complex system including
25
26 bioactive molecules such as antibodies and other proteins in saliva. Although lysozyme
27
28 is present at a high concentration in saliva and possesses a bactericidal activity, it is less
29
30 effective against dental plaque-producing bacteria [33]. Defensins are produced by
31
32 human epithelial cells and active against bacteria, fungi, and enveloped viruses;
33
34 however, their production levels are low [34]. The relation of these molecules to FN has
35
36 not been reported.
37
38

39
40
41 On the other hand, IgA is the most abundant antibody isotype found in the body and
42
43 plays an important role in the immune responses at mucosal surfaces. The main role of
44
45 sIgA antibodies in the oral cavity might be to prevent the colonization of pathogenic
46
47 microorganisms [35]. Secretory IgA does not only prevent adhesion of bacterial cells
48
49 and viruses to mucosal surfaces but also prevent submucosal invasion by stopping
50
51 antigens in the mucosal layer. It also has the effect of eliminating the active enzyme and
52
53 toxin produced by bacteria [35]. It was reported that *F. nucleatum* possessed the ability
54
55 to secrete serine proteases, and the 65 kDa serine protease was found to digest the
56
57
58
59
60

1
2
3 α -chains of immunoglobulin A [36], as the result, the degradation of sIgA may help the
4
5 evasion of the immune system of the host by the bacteria [16]. Therefore, the lower
6
7 levels of sIgA in saliva with a high rate of *F. nucleatum* may lead to reduced protection
8
9 of oral mucosa in addition to the reduced immune system, as the side effect of
10
11 chemotherapy. One possible explanation for chemotherapy-induced FwN is that these
12
13 phenomena led to a decrease in the number of neutrophils as an outcome and the onset
14
15 of fever.
16
17

18
19 Multiple factors are involved in FwN onset, the endpoint of this study. Oral cavity
20
21 immunity and microbial pathogenic virulence are among the predominant oral
22
23 environmental factors of FwN onset. Therefore, we have devised a protocol to measure
24
25 both sIgA and periodontopathic bacteria in this study. The cox-hazard analysis in Table
26
27 3 revealed the significant involvement of sIgA and *F. nucleatum* as independent related
28
29 factors of FwN onset. Therefore, we thought that it was possible to predict FwN risk by
30
31 focusing on sIgA and *F. nucleatum*, as the factor of host defense and attacks by the
32
33 pathogen, respectively. From the results of the ROC curve analysis, *F. nucleatum* / sIgA
34
35 ratio could be applied as predictive indices highly relevant to FwN onset. It was
36
37 demonstrated that some of the items such as age, nutritional status, complications, type
38
39 of cancer, type of anticancer drug and degree of gastrointestinal tract/oral mucosal
40
41 damage, have been shown as risk factors, epidemiologically [14]. In this study, *F.*
42
43 *nucleatum* / sIgA ratio (0.023) was confirmed to be useful as a cut-off line for the
44
45 4-week observation period even if the biases such as age, sex, and serum albumin, were
46
47 considered. These findings may be effective in the future treatment of FwN, in addition
48
49 to the determination of G-CSF prophylactic administration [37], which is currently
50
51 considered to be effective in reducing the incidence rate of FwN.
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 A limitation of this study is the small number of patients studied. It was the inability
4
5 to obtain data regarding the number of oral bacteria and the levels of sIgA in subjects
6
7 with the same type of cancer and/or regimens during the observation periods. Patients
8
9 with acute myelogenous leukemia are generally accepted to most likely develop FwN
10
11 because of the higher intensity of chemotherapies and accompanied severe neutropenia.
12
13 Therefore, we reevaluated and confirmed the importance of the cut-off point, 0.023 of
14
15 the *F. nucleatum* / sIgA ratio, in the prediction of FwN in those excluding myelogenous
16
17 leukemia patients (Supplemental Table 1). However, further validation is needed
18
19 according to cancer types and/or therapeutic regimens.
20
21
22
23

24 In this study, it was possible to evaluate moderate neutropenia (<1,500 / μ L) from the
25
26 medical records of all subjects whereas it was not for severe neutropenia (<500 / μ L).
27
28 However, it is crucial to evaluate the severity of neutropenia regarding the value and the
29
30 duration for the analysis of FN occurrence. Therefore, further study is needed to
31
32 consider deeply the relationship between *F. nucleatum*/ sIgA ratio and FN occurrence by
33
34 adding the data of the severity of neutropenia.
35
36
37

38 Furthermore, oral mucositis during the observation period and other bacteria related
39
40 to oral infection are also required for analysis. It is also necessary to increase the
41
42 number of subjects in future studies, to investigate the involvement of oral bacterial
43
44 infections including *F. nucleatum* in blood, to obtain several kinds of clinical data
45
46 including oral condition during the observation period, and to verify the existence of a
47
48 causal relationship.
49
50
51

52 53 54 55 **Conclusion**

56
57 By investigating the association of chemotherapy-induced FwN, oral bacteria and oral
58
59

1
2
3 immunity, it was revealed that the rate of *F. nucleatum* and the levels of sIgA at baseline
4
5 might be related to FwN onset, and are risk factors. Also, *F. nucleatum* / sIgA ratio
6
7 (<0.023) can be considered as a predictor of FwN onset.
8
9

10 11 12 **Acknowledgments**

13
14 We would like to thank Dr. Kumiko Kagawa, Hirokazu Miki, and Shiro Fujii, medical
15
16 doctors at the Tokushima University Graduate School of Biomedical Sciences, who
17
18 provided valuable support in this study. We also would like to thank Dr. Omar Marianito
19
20 Maningo Rodis, Tokushima University Graduate School of Biomedical Sciences, who
21
22 provided support in amending the manuscript. This study was supported by JSPS
23
24 KAKENHI Grant Number 19K10432 and 19K08839 from the Japan Society for the
25
26 Promotion of Science.
27
28
29
30
31
32
33

34 **Statement of Author Contributions**

35
36 The authors' contributions are as follows: Yuka Sogawa performed all experiments and
37
38 drafted the paper. Makoto Fukui, Shingen Nakamura, Kimiko Sogabe, Ryohei Sumitani,
39
40 and Masami Yoshioka obtained the clinical data from subjects and performed some of
41
42 the experiments. Masahiro Abe contributed to the design and drafting of the paper.
43
44 Daisuke Hinode designed, coordinated the study, and supervised in drafting the paper.
45
46 All authors reviewed the paper critically for content and approved it for submission.
47
48
49
50
51
52

53 **Conflict of interest**

54
55 The authors declare that there is no conflict of interest regarding the publication of this
56
57 paper.
58
59
60
61
62
63
64
65

1
2
3 **References**
4

- 5 1 Klastersky J. Febrile neutropenia. *Curr Opin Oncol.* 1993; 5: 625-33.
6
7 2 Masaoka T. Evidence-based recommendations for antimicrobial use in febrile
8 neutropenia in Japan: executive summary. *Clin Infect Dis.* 2004; 39: S49-52.
9
10 3 de Nauroi J, Novitzky-Basso I, Gill MJ, Marti Marti F, Cullen MH, Roila F.
11 Management of febrile neutropenia: ESMO Clinical Practice Guidelines. *Ann Oncol.*
12 2010; 21(Suppl 5):252-6.
13
14 4 Flowers CR, Seidenfeld J, Bow EJ, Karten C, Gleason C, Hawley DK et al.
15 Antimicrobial prophylaxis and outpatient management of fever and neutropenia in
16 adults treated for malignancy American Society of Clinical Oncology clinical practice
17 guideline: *J Clin Oncol.* 2013; 31: 794-810.
18
19 5 Kasiwazaki H, Matsushita T, Sugita J, Sigematsu A, Kasashi K, Yamazaki Y et al.
20 Professional oral health care reduces oral mucositis and febrile neutropenia in patients
21 treated with allogeneic bone marrow transplantation. *Support Care Cancer.* 2012; 20:
22 367-73.
23
24 6 Terhes G, Piukovics K, Urbán E, Nagy E. Four cases of bacteraemia caused by
25 *Fusobacterium nucleatum* in febrile, neutropenic patients. *J Med Microbiol.* 2011; 60:
26 1046-9.
27
28 7 Harrison T, Bigler L, Tucci M, Pratt L, Malamud F, Thigpen JT et al. Salivary sIgA
29 concentrations and stimulated whole saliva flow rates among women undergoing
30 chemotherapy for breast cancer: an exploratory study. *Spec Care Dentist.* 1998; 18:
31 109-12.
32
33 8 Karolewska E, Konopka T, Pupek M, Chybicka A, Mendak M. Antibacterial
34 potential of saliva in children with leukemia. *Oral Surg Oral Med Oral Pathol Oral*
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

1
2
3 Radiol Endod. 2008; 105: 739-44.
4

5 9 Takamatsu Y. A general description of the clinical guideline for the management of
6 febrile neutropenia. Gan To Kagaku Ryoho. 2013; 40: 697-702. (in Japanese)
7

8
9
10 10 Bradford M. A rapid and sensitive method for the quantification of microgram
11 quantities of protein utilizing the principle of protein-dye binding. Anal Biochem.
12 1976; 72: 248-54.
13
14

15
16
17 11 Moriyama M, Hinode D, Yoshioka M, Sogawa Y, Nishino T, Tangoku A et al. Impact
18 of the use of Kampo medicine in patients with esophageal cancer during
19 chemotherapy: a clinical trial for oral hygiene and oral condition. J Med Invest. 2018;
20 65:184-90.
21
22

23
24
25 12 Yokoyama M, Hinode D, Yoshioka M, Fukui M, Tanabe S, Grenier D et al.
26 Relationship between *Campylobacter rectus* and periodontal status during pregnancy.
27 Oral Microbiol Immunol. 2008; 23: 55-9.
28
29

30
31
32 13 Hajian-Tilaki K. Receiver Operating Characteristic (ROC) Curve Analysis for
33 Medical Diagnostic Test Evaluation. Caspian J Intern Med. 2013; 4: 627-35.
34
35

36
37
38 14 Freifeld AG, Bow EJ, Sepkowitz KA, Boeckh MJ, Ito JI, Mullen CA et al. Clinical
39 practice guideline for the use of antimicrobial agents in neutropenic patients with
40 cancer: 2010 update by the infectious diseases society of America. Clin Infect Dis.
41 2011; 52: e56-93.
42
43

44
45
46 15 Simón-Soro A, Tomás I, Cabrera-Rubio R Catalan MD, Nyvad B, Mira A. Microbial
47 geography of oral cavity. J Dent Res. 2013; 92: 616-21.
48
49

50
51
52 16 Signat B, Roques C, Poulet P, Duffaut D. Role of *Fusobacterium nucleatum* in
53 periodontal health and disease. Curr Issues Mol Biol. 2011; 13: 25-36.
54
55

56
57
58 17 Kolenbrander PE, London J. Adhere today, here tomorrow: oral bacterial adherence.
59
60

- 1
2
3 J Bacteriol. 1993; 175: 3247-52.
4
5 18 Hamada S, Slade HD. Biology, Immunology, and cariogenicity of *Streptococcus*
6
7 *mutans*. Microbiol Rev. 1980; 44: 331-84.
8
9
10 19 Chew J, Zilm PS, Fuss JM, Gully NJ. A proteomic investigation of *Fusobacterium*
11
12 *nucleatum* alkaline-induced biofilms. BMC Microbiol. 2012; 12: 189. doi:
13
14 10.1186/1471-2180-12-189.
15
16
17 20 Gibbons RJ, van Houte J. Bacterial adherence in oral microbial ecology. Annu Rev
18
19 Microbiol. 1975; 29: 19-44.
20
21
22 21. Moritani K, Takeshita T, Shibata Y, Ninomiya T, Kiyohara Y, Yamashita Y.
23
24 Acetaldehyde production by major oral microbes. Oral Dis. 2015; 21: 748–54.
25
26
27 22 Jewett A, Hume WR, Le H, Huynh TN, Han YW, Cheng G et al. Induction of
28
29 apoptotic cell death in peripheral blood mononuclear and polymorphonuclear cells by
30
31 an oral bacterium, *Fusobacterium nucleatum*. Infect Immun. 2000; 68: 1893-8.
32
33
34 23 Tokuyasu H, Harada T, Watanabe E, Okazaki R, Touge H, Kawasaki Y et al.
35
36 Effectiveness of Meropenem for the treatment of aspiration pneumonia in elderly
37
38 patients. Internal Medicine. 2009; 48:129-35.
39
40
41 24 Han YW, Redline RW, Li M, Yin L, Hill GB, McCormick TS. *Fusobacterium*
42
43 *nucleatum* induces premature and term stillbirths in pregnant mice: implication of oral
44
45 bacteria in preterm birth. Infect Immun. 2004; 72: 2272-9.
46
47
48 25 Brennan CA, Garrett WS. *Fusobacterium nucleatum* - symbiont, opportunist and
49
50 oncobacterium. Nat Rev Microbiol. 2019; 17: 156-66.
51
52
53 26 Yamamura K, Baba Y, Nakagawa S, Mima K, Miyake K, Nakamura K et al. Human
54
55 microbiome *Fusobacterium nucleatum* in esophageal cancer tissue is associated with
56
57 prognosis. Clin Cancer Res. 2016; 15: 5574-81.
58
59

- 1
2
3 27 Han YW, Shi W, Huang GT, Kinder Haake S, Park NH, Kuramitsu H et al.
4
5 Interactions between periodontal bacteria and human oral epithelial cells:
6
7 *Fusobacterium nucleatum* adheres to and invades epithelial cells. Infect Immun. 2000;
8
9 68: 3140-6.
10
11
12 28. Han YW. *Fusobacterium nucleatum*: a commensal-turned pathogen. Curr Opin
13
14 Microbiol, 2015; 23: 141-7.
15
16
17 29 Coullioud D, Van der Auwera P, Viot M, Lassent C. Prospective multicentric study of
18
19 the etiology of 1051 bacteremic episodes in 782 cancer patients. CEMIC
20
21 (French-Belgian Study Club of Infectious Diseases in Cancer). Support Care Cancer.
22
23 1993; 1: 34-46.
24
25
26 30 Sogawa Y, Yoshioka M, Fukui M, Nakamura S, Abe M, Hinode D. Effectiveness of
27
28 professional oral health care on oral mucositis in patients undergoing autologous
29
30 hematopoietic stem cell transplant. J Dent Hlth. 2019; 69: 125-30. (in Japanese)
31
32
33 31 Watanabe T, Tobinai K, Shibata T, Tsukasaki K, Morishima Y, Maseki N et al. Phase
34
35 II / III study of R-CHOP-21 versus R-CHOP-14 for untreated indolent B-cell
36
37 non-Hodgkin's lymphoma: JCOG 0203 trial. J Clin Oncol. 2011; 29: 3990-8.
38
39
40 32 Sun H, Chen Y, Zou X, Li Q, Li H, Shu Y et al. Salivary secretory immunoglobulin
41
42 (SIgA) and lysozyme in malignant tumor patients. Biomed Res Int. 2016; 2016:
43
44 8701423. doi: 10.1155/2016/8701423.
45
46
47 33. Wolinsky LE, Caries and cariology, In: Nisengard RJ, Newman MG, editors. Oral
48
49 microbiology and immunology 2nd edition. Philadelphia: Saunders; 1994, p350-1.
50
51
52 34. Cole MF, Lydyard PM. Oral microbiology and the immune response, In: Lamont RJ,
53
54 Burne RA, Lantz MS, Leblanc DJ editors, Oral microbiology and immunology.
55
56 Washington DC: ASM Press; 2006, p205-6.
57
58
59
60
61
62
63
64
65

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

35 Marcott H, Lavoie MC. Oral microbial ecology and the role of salivary immunoglobulin A. *Microbiol Mol Biol Rev.* 1998; 62: 71-109.

36 Bachrach G, Rosen G, Bellalou M, Naor R, Sela MN. Identification of a *Fusobacterium nucleatum* 65 kDa serine protease. *Oral Microbiol Immunol.* 2004; 19: 155-9.

37 Smith TJ, Khatcheressian J, Lyman GH, Ozer H, Armitage JO, Balducci L et al. 2006 update of recommendations for the use of white blood cell growth factors: an evidence-based clinical practice guideline. *J Clin Oncol.* 2006; 24: 3187-3205.

1
2
3 **Figure legends**
4
5
6

7
8 Fig. 1 ROC curve for the rate of *F. nucleatum*, the levels of sIgA and *F. nucleatum* /
9
10 sIgA ratio as a predictor for FwN onset
11
12

13
14 Fig. 2. The Kaplan-Meier curves showing the cumulative hazard of FwN onset
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

Table 1 Type of hematological cancer of subjects in this study

Type	Total subjects (n=32)	
	n	%
Malignant lymphoma	24	75.0
Multiple myeloma	3	9.4
Acute myelogenous leukemia	3	9.4
Chronic myelogenous leukemia	1	3.1
Adult T-cell leukemia/lymphoma	1	3.1

Table 2 Participant characteristics according to the status of FwN onset

Items	Total subjects (n=32)		FwN subjects (n=11)		Non-FwN subjects (n=21)		p-value [§]
	n	%	n	%	n	%	
Gender (male)	19	59.4	7	63.6	12	57.1	1
Regimen							
Anthracycline	27	84.4	10	90.9	17	81.0	0.637
Ifosfamide	1	3.1	0	0.0	1	4.8	1
Cyclophosphamide	25	78.1	8	72.7	17	81.0	0.668
Etoposide	2	6.3	1	9.1	1	4.8	1
Cytarabine	5	15.6	3	33.3	2	9.5	0.31
Antibiotics	27	84.4	11	100	16	76.2	0.138
Diabetes	2	6.3	1	9.1	1	4.8	1
G-CSF	19	59.4	7	63.6	12	57.1	1
Smoking habit (current)	7	21.9	3	33.3	4	19.0	0.668
Items	median	25th, 75th percentile	median	25th, 75th percentile	median	25th, 75th percentile	p-value [¶]
Age (years)	62	52.8, 71.0	53	45.5, 59.5	68	59.0, 73.0	0.014
Albumin (g/dL)	3.9	3.3, 4.0	3.6	3.1, 4.0	3.9	3.5, 4.2	0.116
sIgA (µg/min)	156.7	64.7, 232.9	53.5	30.7, 156.7	190.7	94.6, 269.9	0.026
Protein (mg/min)	0.642	0.410, 1.508	0.515	0.332, 0.831	0.700	0.477, 1.605	0.177
<i>C. rectus</i> (%)	0.024	0.009, 0.055	0.028	0.012, 0.069	0.021	0.006, 0.045	0.692
<i>F. nucleatum</i> (%)	0.405	0.077, 3.204	2.738	0.573, 5.225	0.233	0.042, 1.036	0.088
<i>P. gingivalis</i> (%)	0	0, 0.023	0	0, 0.050	0	0, 0.010	0.646

[§]: Fisher's exact test

[¶]: Mann-Whitney's U test

Table 3 Results from the cox-proportional hazard regression model depicting FwN onset

Covariates		Single variable analysis			Multivariable analysis*		
		HR [†]	95% CI	P-value	HR [†]	95% CI	P-value
Gender	Male [§]						
	Female	0.819	0.240 - 2.798	0.750	2.891	0.238 – 35.096	0.860
Age (year)		0.960	0.928 - 0.993	0.017	0.922	0.913 – 1.040	0.439
Smoking habit	Never or Former [§]						
	Current	1.336	0.354 - 5.042	0.669	0.306	0.041 - 2.264	0.246
Albumin (g/dL)		0.354	0.115 - 1.088	0.070	0.291	0.047 - 1.788	0.183
sIgA (µg/min)		0.992	0.985 - 0.999	0.031	0.984	0.970 - 0.999	0.038
<i>F. nucleatum</i> (%)		1.102	0.970 - 1.253	0.136	1.467	1.071 - 2.008	0.017
Regimen							
Anthracycline	No [§]						
	Yes	1.834	0.235 - 14.343	0.563	0.603	0.025 - 14.578	0.755
Cyclophosphamide	No [§]						
	Yes	0.709	0.188 - 2.673	0.611	1.205	0.077 -18.842	0.894
Cytarabine	No [§]						
	Yes	2.053	0.541 - 7.790	0.291	4.693	0.698 - 31.538	0.112
Diabetes mellitus	No [§]						
	Yes	1.241	0.159 - 9.711	0.837	0.475	0.030 - 7.557	0.598
G-CSF	No [§]						
	Yes	1.241	0.363 – 4.240	0.731	1.046	0.086 - 12.709	0.972

*: Adjusted by gender, age, smoking habit, albumin, *F. nucleatum*, sIgA, diabetes mellitus, G-CSF and regimen.

†: Hazard ration.

§: Reference category.

Table 4 ROC curve analysis regarding FwN onset

Items	AUC †	p-value	cut-off point	Sensitivity	Specificity
sIgA (µg/min)	0.745	0.008	159.446	0.818	0.619
<i>F. nucleatum</i> (%)	0.688	0.069	0.923	0.727	0.714
<i>F. nucleatum</i> / sIgA ratio	0.784	0.003	0.023	0.727	0.792

†: The area under the receiver operating characteristic (ROC) curve.

Table 5 Cox-proportional hazard regression model depicting FwN onset using *F. nucleatum* /sIgA ratio (n=32)

Covariates		Multivariable analysis*		
		HR [†]	95% CI	p-value
Gender	Male [§]			
	Female	1.967	0.273 - 14.194	0.502
Age (year)		0.937	0.869 - 1.011	0.094
Smoking habit	Never or Former [§]			
	Current	0.178	0.018 - 1.742	0.138
Albumin (g/dL)		0.170	0.018 - 1.634	0.125
<i>F. nucleatum</i> / sIgA ratio	<0.023 [§]			
	≥0.023	66.062	3.645 - 1197.37	0.005
Regimen				
Anthracycline	No [§]			
	Yes	0.297	0.009 - 9.964	0.498
Cyclophosphamide	No [§]			
	Yes	0.614	0.573 - 23.608	0.784
Cytarabine	No [§]			
	Yes	3.679	0.020 - 3.681	0.170
Diabetes mellitus	No [§]			
	Yes	0.273	0.326 - 118.327	0.328
G-CSF	No [§]			
	Yes	6.215	0.100 - 118.967	0.224

*: Adjusted by gender, age, smoking habit, albumin, [*F. nucleatum*/ sIgA ratio], diabetes mellitus, G-CSF and regimen.

†: Hazard ration.

§: Reference category.



