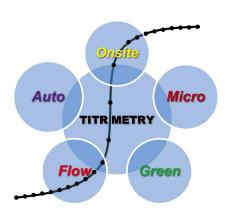
Highlights

Titrimetry

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Titrimetry is among absolute methods, that rely upon accurately known fundamental constants for calculating the amount of an analyte. The origin of this technique goes back to Geffroy in 1729; he determined the concentration of vinegar by noting the amount of solid potassium carbonate that could be added before effervescence stopped.1 Titrimetry was developed in the eighteenth century, but is still widely used today due to its high precision and versatility. For example, Masadome et al. reported a photometric colloidal titration method of polyhexamethylene biguanide hydrochloride, which is used for disinfectants in personal-care products, using crystal violet as a color indicator.² Zhi et al. applied the Karl Fischer coulometric titration method to determine the water content of nitrogen-containing hydrogen sulfide.3 Anderson et al. utilized a potentiometric titration curve to calculate the concentration of polyquaternium polymers which are used for medical studies, wastewater treatment, and environmental contaminant remediation.4 Routine tests of the chemical oxygen demand (COD) is mostly achieved by batchwise titration, but in recent years several alternative methods have been reported. Chen et al. proposed a gas-phase molecular absorption spectrometry-based COD analyzer.⁵ The proposed method improved the analysis speed, efficiency, accuracy, and stability compared to the traditional batch-wise titration. Hue et al. reported a 3-step chemiluminescence method for COD determination, i.e., i) treatment of a sample with permanganate under heating, ii) treatment of excess permanganate with pyrogallol, and iii) measurement of excess pyrogallol by chemiluminescence reaction with permanganate.⁶ This method was continuous, sensitive, and low cost compared to the conventional titration method. To achieve green analytical chemistry, many techniques have been integrated into titrimetry. Kanna et al. proposed reusable solid-phase sorbed indicators for



Evolution of titrimetry.

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solids, using anion exchange resins and kaolin clay as supports, demonstrated promising results for determining acidity in weak acid samples. Paengnakorn et al. constructed a sequential injection analysis-lab-at-valve system for automated micro-scale titration.8 This system readily reduced the volume of sample/ reagents on only a micro-liter scale, significantly lower than that used in conventional batch titration. Ochiai et al. studied a feedback-based and subsequent fixed triangular wave-controlled flow ratiometry.9 They introduced air-segmentation to suppress axial dispersion in flow titration, and found that the applicable range was extended especially to lower titrand concentrations. Kawakubo et al. designed a microtitration system based on the counting of titrant droplets for precise on-site analysis. 10 On-site measurements of tap water collected in an office kitchen were performed, and the analytical results agreed with those of conventional titration with an error of 3%.

reducing reagents used in titrimetry.7 The indicator-sorbed-

Titrimetry, developed about 300 years ago, has never lost its importance despite the emergence of various new analytical methods. This classical analytical method will continue to evolve and develop with changing the needs of the times.

Keywords Titrimetry, automated analysis, on-site analysis, flow analysis, green chemistry

References

- G. D. Christian, P. K. Dasgupta, and H. A. Schung, "Analytical Chemistry", 7th ed., 2013, Wiley Global Education.
- 2. T. Masadome, T. Miyanishi, K. Watanabe, H. Ueda, and T. Hattori, *Anal. Sci.*, **2011**, *27*, 817.
- X. Zhi, H. Wang, B. Liu, X. Song, Z. Li, and J. Li, *Anal. Sci.*, 2019, 35, 777.
- E. L. Anderson, P. D. Samaniego, and P. Bühlmann, *Anal. Sci.*, 2019, 35, 679.
- X. Chen, L. Peng, J. Wang, D. Zhang, Y. Zhao, Q. Zhao, and T. Li, *Anal. Sci.*, 2020, 36, 841.
- D. T. K. Hue, S. Hashimoto, H. Nishikawa, Y. Maeda, and N. Takenaka, *Anal. Sci.*, 2017, 33, 931.
- 7. M. Kanna, S. Somnam, W. Wongwilai, and K. Grudpan, *Anal. Sci.*, **2019**, *35*, 347.
- 8. P. Paengnakorn, S. Chanpaka, K. Watla-Iad, W. Wongwilai, and K. Grudpan, *Anal. Sci.*, **2019**, *35*, 219.
- 9. J. Ochiai, S. Oka, T. Hirasaka, E. Tomiyama, H. Kubo, K. Okamoto, M. Takeuchi, and H. Tanaka, *Anal. Sci.*, **2020**, *36*, 703.
- S. Kawakubo, T. Omori, Y. Suzuki, and I. Ueta, *Anal. Sci.*, 2018, 34, 243.