

# Population-Based Impact of Smoking, Drinking, and Genetic Factors on HDL-cholesterol Levels in J-MICC Study Participants

Yora Nindita<sup>1,2</sup>, Masahiro Nakatochi<sup>3</sup>, Rie Ibusuki<sup>1</sup>, Ippei Shimoshikiryō<sup>1</sup>, Daisaku Nishimoto<sup>1,4</sup>, Keiichi Shimatani<sup>5</sup>, Toshiro Takezaki<sup>1</sup>, Hiroaki Ikezaki<sup>6,7</sup>, Masayuki Murata<sup>7</sup>, Megumi Hara<sup>8</sup>, Yuichiro Nishida<sup>8</sup>, Takashi Tamura<sup>9</sup>, Asahi Hishida<sup>9</sup>, Mako Nagayoshi<sup>9</sup>, Rieko Okada<sup>9</sup>, Keitaro Matsuo<sup>10</sup>, Hidemi Ito<sup>11</sup>, Haruo Mikami<sup>12</sup>, Yohko Nakamura<sup>12</sup>, Takahiro Otani<sup>13</sup>, Sadao Suzuki<sup>13</sup>, Teruhide Koyama<sup>14</sup>, Etsuko Ozaki<sup>14</sup>, Kiyonori Kuriki<sup>15</sup>, Naoyuki Takashima<sup>16,17</sup>, Naoko Miyagawa<sup>17,18</sup>, Kokichi Arisawa<sup>19</sup>, Sakurako Katsuura-Kamano<sup>19</sup>, Yukihide Momozawa<sup>20</sup>, Michiaki Kubo<sup>20</sup>, Kenji Takeuchi<sup>9</sup>, and Kenji Wakai<sup>9</sup>,  
for the Japan Multi-Institutional Collaborative Cohort Study Group

<sup>1</sup>Department of International Island and Community Medicine, Kagoshima University Graduate School of Medical and Dental Sciences, Kagoshima, Japan

<sup>2</sup>Department of Pharmacology and Therapeutic, Faculty of Medicine, Diponegoro University, Semarang, Indonesia

<sup>3</sup>Public Health Informatics Unit, Department of Integrated Health Sciences, Nagoya University Graduate School of Medicine, Nagoya, Japan

<sup>4</sup>School of Health Sciences, Faculty of Medicine, Kagoshima University, Kagoshima, Japan

<sup>5</sup>Division of Nursing, Higashigaoka Faculty of Nursing, Tokyo Healthcare University, Tokyo, Japan

<sup>6</sup>Department of Comprehensive General Internal Medicine, Kyushu University Graduate School of Medical Sciences, Faculty of Medical Sciences, Fukuoka, Japan

<sup>7</sup>Department of General Internal Medicine, Kyushu University Hospital, Fukuoka, Japan

<sup>8</sup>Department of Preventive Medicine, Faculty of Medicine, Saga University, Saga, Japan

<sup>9</sup>Department of Preventive Medicine, Nagoya University Graduate School of Medicine, Nagoya, Japan

<sup>10</sup>Division of Cancer Epidemiology and Prevention, Aichi Cancer Center Research Institute, Nagoya, Japan

<sup>11</sup>Division of Cancer Information and Control, Aichi Cancer Center Research Institute, Nagoya, Japan

<sup>12</sup>Cancer Prevention Center, Chiba Cancer Center Research Institute, Chiba, Japan

<sup>13</sup>Department of Public Health, Nagoya City University Graduate School of Medical Sciences, Nagoya, Japan

<sup>14</sup>Department of Epidemiology for Community Health and Medicine, Kyoto Prefectural University of Medicine, Kyoto, Japan

<sup>15</sup>Laboratory of Public Health, Division of Nutritional Sciences, School of Food and Nutritional Sciences, University of Shizuoka, Shizuoka, Japan

<sup>16</sup>Department of Public Health, Kindai University Faculty of Medicine, Osaka, Japan

<sup>17</sup>Department of Public Health, Shiga University of Medical Science, Otsu, Japan

<sup>18</sup>Department of Preventive Medicine and Public Health, Keio University School of Medicine, Tokyo, Japan

<sup>19</sup>Department of Preventive Medicine, Tokushima University Graduate School of Biomedical Sciences, Tokushima, Japan

<sup>20</sup>Laboratory for Genotyping Development, Center for Integrative Medical Sciences, RIKEN, Yokohama, Japan

Received March 26, 2021; accepted August 2, 2021; released online August 21, 2021

## ABSTRACT

**Background:** Environmental and genetic factors are suggested to exhibit factor-based association with HDL-cholesterol (HDL-C) levels. However, the population-based effects of environmental and genetic factors have not been compared clearly. We conducted a cross-sectional study using data from the Japan Multi-Institutional Collaborative Cohort (J-MICC) Study to evaluate the population-based impact of smoking, drinking, and genetic factors on low HDL-C.

**Methods:** Data from 11,498 men and women aged 35–69 years were collected for a genome-wide association study (GWAS). Sixty-five HDL-C-related SNPs with genome-wide significance ( $P < 5 \times 10^{-8}$ ) were selected from the GWAS catalog, of which seven representative SNPs were defined, and the population-based impact was estimated using population attributable fraction (PAF).

**Results:** We found that smoking, drinking, daily activity, habitual exercise, egg intake, BMI, age, sex, and the SNPs *CETP* rs3764261, *APOA5* rs662799, *LIPC* rs1800588, *LPL* rs328, *ABCA1* rs2575876, *LIPG* rs3786247, and *APOE* rs429358 were associated with HDL-C levels. The gene-environmental interactions on smoking and drinking were not statistically significant. The PAF for low HDL-C was the highest in men (63.2%) and in rs3764261 (31.5%) of the genetic factors, and the PAFs of smoking and drinking were 23.1% and 41.8%, respectively.

**Conclusion:** The present study showed that the population-based impact of genomic factor *CETP* rs3764261 for low HDL-C was higher than that of smoking and lower than that of drinking.

**Key words:** HDL-cholesterol; drinking; smoking; single nucleotide polymorphism; gene-environmental interaction

Copyright © 2021 Yora Nindita et al. This is an open access article distributed under the terms of Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Address for correspondence. Toshiro Takezaki, MD, DMSc, Department of International Island and Community Medicine, Kagoshima University Graduate School of Medical and Dental Sciences, 8-35-1 Sakuragaoka, Kagoshima 890-8544, Japan (e-mail: takezaki@m.kufm.kagoshima-u.ac.jp).

## INTRODUCTION

Low serum levels of HDL-cholesterol (HDL-C) are associated with an increased risk of cardiovascular disease (CVD).<sup>1,2</sup> As clinically available drugs that can enhance HDL-C levels are limited, genetic and environmental factors play an important role in the alleviation of CVD risk. Smoking, alcohol intake, physical activity, BMI, and diet intake have been confirmed to be environmental factors that affect HDL-C levels.<sup>3–6</sup>

The effects of genetic factors, such as single nucleotide polymorphism (SNPs) in various enzymes-encoding genes, on HDL-C levels have also been reported.<sup>7</sup> Although the regulation of HDL-C metabolism is a complex process, enzymes in the reverse cholesterol transport (RCT) system, such as ABCA1, LCAT, cholesteryl ester transfer protein (CETP), hepatic lipase (LIPC), APOA1/C3/A4/A5, scavenger receptor class B type I (SCARB1), and LPL, play a major role in it.<sup>2</sup> Multiple SNPs have been reported to be associated with HDL-C levels, and among the genes harboring such SNPs, the genetic variants of *CETP* have been observed to exert a greater influence on HDL-C levels.<sup>8–11</sup> Furthermore, besides association with SNPs in RCT-related genes, the association with several other SNPs, such as those in genes encoding endothelial lipase (LIPG) and APOE, which are related to lipoprotein dynamism, has been reported.<sup>10,12</sup>

The majority of studies on environmental and genetic factors that affect HDL-C levels focus on factor-based association with respect to individual risk and susceptibility, and the population-based impact of environmental and genetic factors on HDL-C levels has not been clearly evaluated. The population-based impact of a factor is an important aspect for public health. The population-based impact of various environmental factors on HDL-C levels can be estimated based on the impact of the association and prevalence of each factor. However, the population-based impact of genetic factors is difficult to estimate, because several SNPs are detected in each enzyme-encoding gene; the impact of the association of each SNP with HDL-C levels will differ, and the prevalence of the allele containing each SNPs will differ as well. Therefore, studies that investigate the combined effect of HDL-C-related SNPs limit their assessment to certain representative SNPs.<sup>9</sup> Furthermore, gene-environment interaction may influence HDL-C levels as well.<sup>13,14</sup>

Among environmental factors, smoking and drinking habits significantly affect the reduction or increase in HDL-C levels, respectively.<sup>2,9,15</sup> These factors are suitable candidates for the estimation of the population-based impact of environmental factors on HDL-C levels, while also taking into account the interaction with genetic factors. In such cases, GWAS are suitable for evaluating the overall scenario. GWAS on the effects of HDL-C-related SNPs on ethnic populations, including the Japanese population, have been performed earlier, and all HDL-C-related SNPs have been listed in the catalog.<sup>16,17</sup>

To investigate the population-based impact of smoking, drinking, and genetic factors on low HDL-C, we conducted a relatively large-sized cross-sectional study using data on environmental factors and GWAS from the Japan Multi-Institutional Collaborative Cohort (J-MICC) Study.

## METHODS

### Study population

The J-MICC Study was a large-scaled cohort study that

commenced in 2005; it investigated the host- and environment-related factors that affect cancer and other lifestyle-related diseases.<sup>18–20</sup> In brief, data on the lifestyles and medical history of patients were collected using questionnaires, while blood samples and health checkup results were collected during the baseline survey after written informed consent was obtained. The participants were recruited from among health-checkup examinees by the local government, private companies, and health checkup centers; responders who posted responses to regional residents and first-visit outpatients at cancer center. The subjects ( $n = 14,555$ ) of the GWAS selected from among the J-MICC Study participants were aged from 35–69 years and belonged to 11 prefectures of Japan (Chiba, Shizuoka, Aichi, Shiga, Kyoto, Tokushima, Fukuoka, Saga, Nagasaki, Kagoshima, and Okinawa); participants were selected by ten research institutes and universities. The present study excluded data that did not include information on HDL-C levels (all participants [ $n = 2,296$ ] from the Chiba study region and the Aichi Cancer Center and some participants [ $n = 187$ ] from other institutes), smoking ( $n = 180$ ), and drinking ( $n = 24$ ); and from cases of withdrawal ( $n = 21$ ). Data from certain subjects qualified for multiple exclusion criteria. The final number of eligible subjects was 11,498 (the dataset used in the present study was decided upon on March 12, 2020, version 20200312).

The ethics committees of Nagoya University Graduate School of Medicine, Kagoshima University Graduate School of Medical and Dental Sciences, and other participating institutes and universities approved the protocol.

### Questionnaire survey

A standardized structured questionnaire was used in the J-MICC Study to collect information regarding lifestyle factors and medical history of the subjects.<sup>19</sup> The questionnaire was evaluated by trained staff to ensure completeness and consistency.

### HDL-C level assessment

Venous blood samples were collected from the subjects in sitting position during a period of fasting. The mean duration of fasting was 9.8 h. The blood samples were separated into serum, plasma, and buffy coat fractions, and stored directly at  $-80^{\circ}\text{C}$  on the day of sampling. The serum HDL-C levels were measured at the respective institutes for health checkup or medical examination in each study region.<sup>21</sup>

### Quality of samples and SNPs during genotyping

DNA was extracted from the buffy coat fractions using a BioRobot M48 Workstation (Qiagen Group, Tokyo, Japan) at Nagoya University, using samples from all regions except Fukuoka and KOPS (Kyushu and Okinawa Population Study); DNA was extracted from the samples from these two regions at Kyushu University using an automatic nucleic acid isolation system (NA-3000; Kurabo, Co., Ltd, Osaka, Japan). Next, the DNA samples were genotyped at the RIKEN Center for Integrative Medicine using a HumanOmniExpressExome-8 v1.2 BeadChip array (Illumina Inc., San Diego, CA, USA). The number of low-quality DNA samples was 463, which were excluded from the analysis. The subjects for whom sex information in the questionnaire was inconsistent with that revealed by the genotyping results were excluded. Furthermore, the identity-by-descent method implemented in the PLINK 1.9 software<sup>22</sup> was used to identify close relationship pairs (pi-hat

>0.1875) and the sample from each pair was excluded. The subjects ( $n = 34$ ) with non-Japanese estimated ancestries<sup>23</sup> were also excluded by principal component analysis (PCA)<sup>24</sup> using a 1,000 Genomes reference panel (phase 3).<sup>25</sup>

SNPs with a genotype call rate  $<0.98$ , a Hardy-Weinberg equilibrium exact test  $P$ -value  $<1 \times 10^{-6}$ , and a low minor allele frequency (MAF)  $<0.01$ , or a departure from the allele frequency computed from the 1,000 Genomes Phase 3 EAS (East Asian) samples; and non-autosomal SNPs were excluded. Such quality control filtering resulted in 14,091 individuals and 570,162 SNPs.

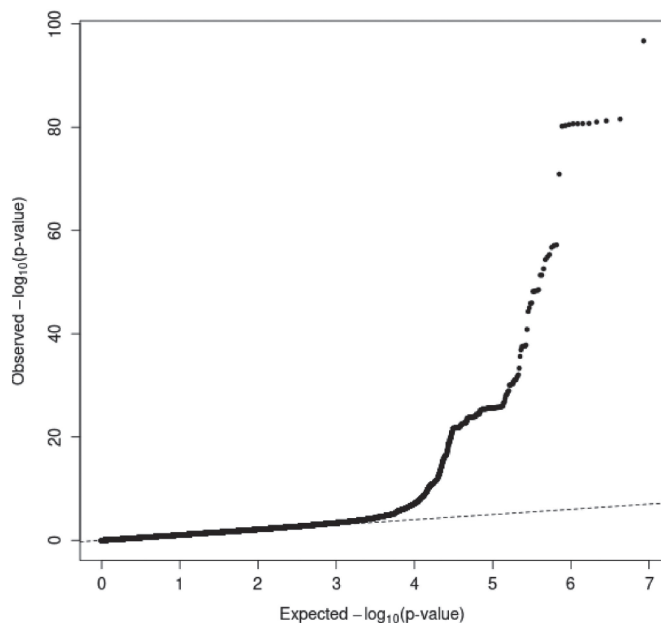
### Genotype imputation and post-imputation quality control

The imputation of genotypes in autosomal chromosomes was performed using SHAPEIT2<sup>26</sup> and Minimac3<sup>27</sup> software with the 1,000 Genomes reference panel (phase 3).<sup>25</sup> The imputation procedure displayed 47,109,431 SNPs from 570,162 SNPs.

The SNPs with imputation quality  $r^2 < 0.3$  were excluded in the post-imputation quality control step. The number of eligible SNPs was 12,617,547.

### Selection of HDL-C-related SNPs

On August 27, 2019, HDL-C-related SNPs were systematically selected from the GWAS catalog (<https://www.ebi.ac.uk/gwas/>) (the database of published GWAS), which included 499 SNPs from all ethnic population.<sup>16,17</sup> Next, 65 SNPs among these were selected for the present study, which had  $P$ -values of genome-wide significance ( $P < 5 \times 10^{-8}$ ) in the present analysis (eTable 1). The Q-Q plot showed the apparently different distribution of the present observed  $\log_{10}(P\text{-value})$  of the 65 SNPs against the expected  $\log_{10}(P\text{-value})$  (Figure 1). Although the association for rs921919 in *SCARB1* (12q24.31) indicated



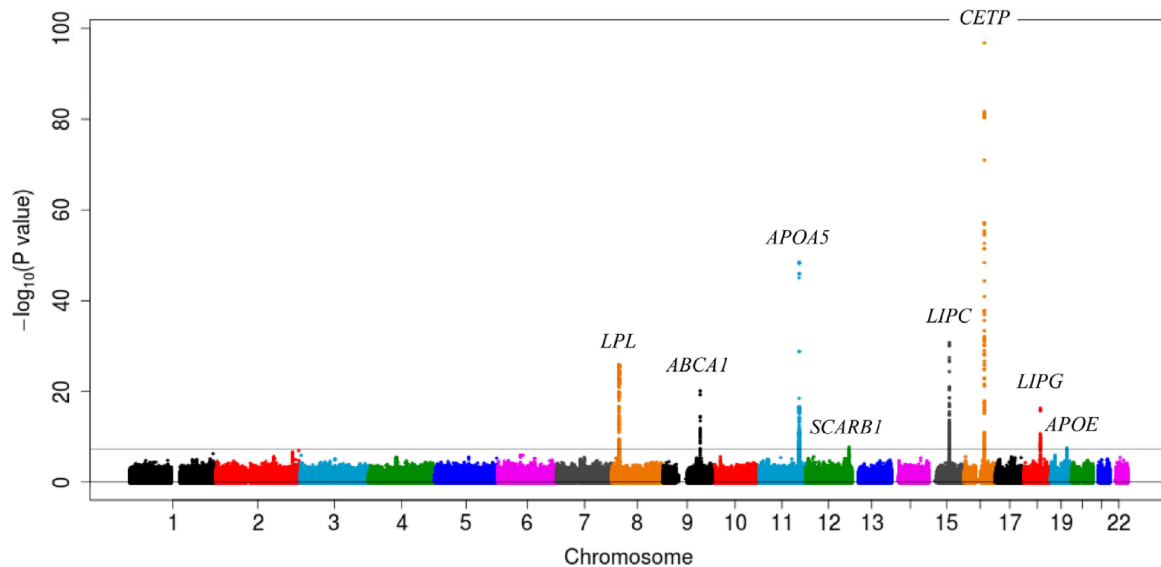
**Figure 1.** Q-Q plot for  $P$  values from original GWAS data. The vertical and horizontal axes indicate observed and expected  $-\log_{10}(P\text{ value})$  for tests of association between SNPs and HDL-C, respectively. GWAS, genome-wide association study; HDL-C, high-density lipoprotein cholesterol; SNP, single-nucleotide polymorphism.

genome-wide significance, this was not included in the present analysis because this SNP was not previously reported to be associated with HDL-C levels and were not listed in the GWAS catalog. Other SNPs in *SCARB1* listed in the GWAS catalog were not genome-wide significant in the present analysis.

### Statistical analysis

The subjects were divided into two categories based on the smoking status (“never” and “former”  $\geq 1$  year) vs “current” [include smokers within 1 year after quitting]), because the HDL-C levels apparently differed between subjects with the “current” and “never” statuses, and with respect to the duration after quitting. The subjects were also divided into two categories based on the drinking status (non-, former, and current moderate drinkers [ $<20$  g/day] vs current heavy drinkers  $\geq 20$  g/day), as the Japanese Ministry of Health, Labour and Welfare recommends alcohol intake in moderation (at  $<20$  g/day); the HDL-C levels apparently differed between the two categories.<sup>28</sup> The duration and intensity of daily activity (hard work and walking) and the frequency and intensity of habitual exercise were used to estimate the metabolic equivalents (METs). The estimation of METs-hour per day was based on the duration and intensity of exercise, with 3.0 for walking, 3.3 for light exercise, 4.0 for moderate exercise, 4.5 for heavy work, and 8.0 for heavy exercise.<sup>29</sup> Daily activity was classified as  $<8.25$  METs-h/day and  $\geq 8.25$  METs-h/day at the median value. Habitual exercise was classified as  $<0.728$  METs-h/day and  $\geq 0.728$  METs-h/day at the median value. Egg intake was selected as a representative HDL-C-related dietary factor.<sup>2,9</sup> There were two categories for BMI with comparable number of male and female subjects in each. The association between HDL-C levels (continuous) and non-genetic factors, such as smoking and drinking habits, was tested using multivariate linear regression analysis after adjusting for the following HDL-C-related factors: age ( $<57$  vs  $\geq 57$  years), sex, smoking, drinking, daily activity, habitual exercise, egg intake, and BMI. Dummy variables of 0 and 1 were used for all independent variables. Statistical analyses for non-genetic factors were performed using Stata software (version 12; Stata Corp., College Station, TX, USA), and differences with  $P$ -value  $<0.05$  were considered statistically significant.

The selected HDL-C-related 65 SNPs were divided into seven categories based on the gene and cytoBand groups (eTable 1). The Manhattan plot for total SNPs in the present GWAS consistently showed seven peaks with genome-wide significance, with the exception of a single peak corresponding to rs921919 in *SCARB1* with genome-wide significance yet unlisted in the GWAS catalog (Figure 2). Next, the seven SNPs with the highest coefficients and lowest  $P$ -values from each of the seven groups were selected. The association between HDL-C levels (continuous) and genetic factors, and the interaction were tested using multivariate linear regression analysis in epacts v3.2.6 software (<https://genome.sph.umich.edu/wiki/EPACTS>), after adjusting for the HDL-C-related factors and first five principal components. Dummy variables of 0, 0.5, and 1 were used for the number of alternative alleles (0, 1, and 2) as independent variables in order to compare the impact of coefficients on non-genetic factors (dummy variables of 0 and 1), and the coefficients and 95% confidence intervals (CIs) were estimated. Differences with  $\alpha = 5 \times 10^{-8}$  were considered statistically significant in the GWAS. We applied the Bonferroni correction ( $P < 0.00077$ ) for evaluating the differences in interaction of smoking or drinking



**Figure 2.** Manhattan plot ( $-\log_{10}$  of the  $P$  value based on genomic location) of the association between the SNPs denoted in the original GWAS and the HDL-C levels shows the formation of eight peaks over the line representing  $P < 5 \times 10^{-8}$  for *LPL* (8p21.3), *ABCA1* (9q31.1), *APOA5* (11q23.3), *SCARB1* (12q24.31), *LIPC* (15q21.3), *CETP* (16q13), *LIPG* (18q21.1), and *APOE* (19q13.32). The horizontal line represents the genome-wide significance level ( $\alpha = 5 \times 10^{-8}$ ). GWAS, genome-wide association study; HDL-C, high-density lipoprotein cholesterol; SNP, single-nucleotide polymorphism.

with the 65 SNPs to reduce the chances of introducing an alpha error by multiple hypothesis testing.

The population-based impact of the non-genetic and genetic factors was estimated using population attributable fraction (PAF).<sup>30,31</sup> First, the odds ratio (OR) for low HDL-C (<40 mg/dL) was estimated, and the PAF was calculated as;

$$PAF = P \times \frac{(OR - 1)}{OR} \times 100 (\%)$$

where  $P$  is the proportion of the exposure in subjects with low HDL-C. The reference exposure group was defined as those with the minimum risk for low HDL-C, ie smoking habit (“never” and “former” [ $\geq 1$  year]), drinking habit ( $\geq 20$  gram alcohol/day), daily activity ( $\geq 8.25$  METs/day), habitual exercise ( $\geq 0.73$  METs/day), egg intake ( $\geq 3$  times/week), BMI (<23.0 kg/m<sup>2</sup>), age (<57 years), and sex (women) in the non-genetic factors; and rs3764261, rs662799, rs1800588, rs328, and rs3786247 (referent and alternative allele hetero-genotype, and alternative allele homo-genotype), as well as rs2575876 and rs429358 (referent allele homo-genotype), in the genetic factors. Dummy variables of 0 and 1 were used for both the non-genetic and genetic factors. When the PAF of the combined SNPs was estimated, the accumulation in 6 SNPs was categorized according to the number of the high-risk genotypes for low HDL-C by individual regardless kind of SNPs (ie, 0–1 SNPs for reference, 2 SNPs, 3 SNPs and 4–6 SNPs). The SNP of rs1800588 was excluded from this accumulation analysis, because the OR for low HDL-C was not statistically significant. The ORs and their 95% CIs were estimated using logistic model after adjusting for age, sex, smoking, drinking, daily activity, habitual exercise, egg intake, and BMI.

## RESULTS

The distribution pattern of male and female subjects in the two age groups (35–56 years and 57–69 years) was almost similar

(Table 1). The prevalence of current smokers was 34.9% among male and 7.3% among female subjects (19.7% in both), and that of heavy drinkers was 42.7% in males and 5.1% in females

**Table 1.** Age-, environmental factor-, BMI-, and HDL-C level-based distribution of study subjects divided by sex

	Number (%)	
	Men	Women
Age, years		
35–56	2,595 (50.7)	3,280 (52.4)
57–69	2,519 (49.3)	2,976 (47.6)
Total	5,114 (100)	6,256 (100)
Smoking		
Never and former ( $\geq 1$ year) smokers	3,329 (65.1)	5,802 (92.7)
Current smokers <sup>a</sup>	1,785 (34.9)	454 (7.3)
Drinking		
Non-, former and moderate drinkers <sup>b</sup>	2,933 (57.4)	5,936 (94.9)
Heavy drinkers <sup>c</sup>	2,181 (42.7)	320 (5.1)
Daily activity		
<8.25 METs-h/day	3,102 (60.7)	3,484 (55.7)
$\geq 8.25$ METs-h/day	2,012 (39.3)	2,772 (44.3)
Habitual exercise		
<0.73 METs-h/day	2,447 (47.9)	3,268 (52.2)
$\geq 0.73$ METs-h/day	2,667 (52.2)	2,988 (47.8)
Egg intake		
<3 times/week	3,659 (71.6)	4,377 (70.0)
$\geq 3$ times/week	1,455 (28.5)	1,879 (30.0)
BMI, kg/m <sup>2</sup>		
<23	2,066 (40.4)	3,789 (60.6)
$\geq 23$	3,048 (59.6)	2,467 (39.4)
HDL-C		
<40 mg/dL	454 (8.9)	111 (1.8)
$\geq 40$ mg/dL	4,660 (91.1)	6,145 (98.2)

BMI, body mass index; HDL-C, high-density lipoprotein cholesterol; METs, metabolic equivalents.

<sup>a</sup>Smokers within 1 year after quitting were included.

<sup>b</sup><20 g alcohol/day.

<sup>c</sup> $\geq 20$  g alcohol/day.

**Table 2.** Association between HDL-C levels and environmental factors determined in multivariate regression analysis

	Coeff. <sup>a</sup>	95% CI	P-value
Smoking (current)	-5.407	-6.133 to -4.681	<0.001
Drinking (≥20 g alcohol/day)	7.274	6.549 to 8.000	<0.001
Daily activity (≥8.25 METs-h/day)	1.033	0.493 to 1.574	<0.001
Habitual exercise (≥0.73 METs-h/day)	1.480	0.938 to 2.021	<0.001
Egg intake (≥3 times/week)	0.856	0.270 to 1.442	0.004
BMI (≥23.0 kg/m <sup>2</sup> )	-8.738	-9.283 to -8.194	<0.001
Age (≥57 years)	-1.496	-2.040 to -0.952	<0.001
Sex (women)	9.567	8.931 to 10.203	<0.001

BMI, body mass index; CI, confidence interval; Coeff., coefficient; HDL-C, high density lipoprotein cholesterol; METs, metabolic equivalents.

<sup>a</sup>Adjusted for age, sex, smoking, drinking, daily activity, habitual exercise, egg intake, and BMI. The coefficient value represents change in HDL-C per dummy variable (0, 1) of environmental factors.

(22.0% in both). The prevalence of low HDL-C (<40 mg/dL) was 8.9% in male and 1.8% in female subjects (5.0% in both sexes).

Drinking ( $P < 0.001$ ), daily activity ( $P < 0.001$ ), habitual exercise ( $P < 0.001$ ), egg intake ( $P = 0.004$ ), and sex ( $P < 0.001$ ) were associated positively with the HDL-C levels, while smoking ( $P < 0.001$ ), BMI ( $P < 0.001$ ), and age ( $P < 0.001$ ) were associated negatively (Table 2).

The seven major SNPs selected from the 65 SNPs in the GWAS catalog according to the gene and cytoBand groups were rs3764261 in *HERPUD1-CETP* (16q13), rs662799 in *APOA5* (11q23.3), rs1800588 in *LIPC* (15q21.3), rs328 in *LPL* (8p21.3), rs2575876 in *ABCA1* (9q31.1), rs3786247 in *LIPG* (18q21.1), and rs429358 in *APOE* (19q13.32) (Table 3). The frequencies (0.100 to 0.649) and coefficients (-4.003 to 8.863) varied for each SNP, and the highest coefficient was observed for rs3764261.

The HDL-C levels varied for each genotype group based on the smoking and drinking status (Table 4). The highest HDL-C level (mean 74.6; 95% CI, 70.8–78.4 mg/dL) was observed in heavy drinkers with the rs3764261 alternative homo-genotype (AA), while the lowest was observed in current smokers with the rs662799 referent homo-genotype (GG) and hetero-genotype (GA). The gene-environment interactions between the seven SNPs and smoking/drinking were not statistically significant, and the lowest  $P$ -value of 0.004 was higher than the  $P$ -value obtained after applying Bonferroni correction ( $P < 0.00077$ ). These interactions were not statistically significant for all 65 SNPs selected from the GWAS catalog (eTable 1). No significant interaction was observed in the subgroup analysis based on sex (data not shown in eTable 1).

**Table 3.** Multivariate regression analysis between HDL-C levels and seven HDL-C related SNPs from the GWAS catalog

SNP	cytoBand	REF/ALT	Gene	Frequency of ALT	Coeff. <sup>a</sup>	95% CI	P-value
rs3764261	16q13	C/A	<i>HERPUD1, CETP</i>	0.207	8.863	7.958 to 9.770	$6.07 \times 10^{-82}$
rs662799	11q23.3	G/A	<i>APOA5</i>	0.649	5.713	4.932 to 6.494	$1.12 \times 10^{-46}$
rs1800588	15q21.3	C/T	<i>LIPC</i>	0.510	4.447	3.700 to 5.194	$1.76 \times 10^{-31}$
rs328	8p21.3	C/G	<i>LPL</i>	0.126	6.136	5.006 to 7.266	$1.77 \times 10^{-26}$
rs2575876	9q31.1	G/A	<i>ABCA1</i>	0.276	-4.003	-4.840 to -3.164	$7.67 \times 10^{-21}$
rs3786247	18q21.1	T/G	<i>LIPG</i>	0.460	3.209	2.452 to 3.966	$1.02 \times 10^{-16}$
rs429358	19q13.32	T/C	<i>APOE</i>	0.100	-3.594	-4.864 to -2.322	$2.89 \times 10^{-8}$

ALT, alternative allele; BMI, body mass index; CI, confidence interval; Coeff., coefficient; GWAS, genome-wide association study; HDL-C, high density lipoprotein cholesterol; REF, referent allele; SNP, single nucleotide polymorphism.

<sup>a</sup>Adjusted for age, sex, smoking, drinking, daily activity, habitual exercise, egg intake and BMI. The coefficient value represents change in HDL-C per ALT allele copy (0, 1, 2) for the SNP.

The ORs for low HDL-C were statistically significant for several non-genetic factors, including smoking, drinking, BMI, age and sex, and for the genetic factors, and six of the seven SNPs (except rs1800588) (Table 5). The PAF for low HDL-C in the non-genetic factors was the highest in men (63.2%), and the PAFs of smoking and drinking were 23.1% and 41.8%, respectively. The PAF for low HDL-C in the genetic factors was the highest in rs3764261 (31.5%), which was higher than that of smoking and lower than that of drinking. The impact of the PAFs of three SNPs (25.5%) and 4–6 SNPs (23.7%) according to the number of SNPs with high-risk genotype for low HDL-C was similar to that of smoking, although the ORs for low HDL-C showed an apparent increasing trend with the number of SNPs with higher-risk genotype ( $P < 0.001$ ).

## DISCUSSION

In the present study, we observed significant associations between HDL-C levels and smoking, drinking, daily activity, habitual exercise, egg intake, BMI, age, sex, and seven SNPs in *CETP*, *APOA5*, *LIPC*, *LPL*, *ABCA1*, *LIPG*, and *APOE*. The PAFs, as a population-based impact, for low HDL-C were the highest in men on the non-genetic factors and in *CETP* rs3764261 on the genetic factors. The impact of the genetic factor PAF was higher than that of smoking and was lower than that of drinking.

Genetic factors that affect HDL-C levels, such as SNPs, are primarily associated with genes that encode enzymes from the RCT system, such as *ABCA1*, *LCAT*, *CETP*, *LIPC*, *APOA1/C3/A4/A5*, *SCARB1*, and *LPL*.<sup>2,7</sup> The SNPs in the corresponding genes, except those in *LCAT* and *SCARB1*, were considered among the seven major SNPs selected in the present analysis. The SNPs in *SCARB1* were not included because the two SNPs with genome-wide significance were not listed in the GWAS catalog, and the lowest  $P$ -value for the *SCARB1* SNP (rs838886) listed in the catalog was higher than the genome-wide significance ( $P = 7.34 \times 10^{-8}$ ; data not shown in eTable 1). As the MAF of *LCAT* was less than 0.01, the SNPs of *LCAT* were excluded from the GWAS analysis. The SNPs in *LIPG* and *APOE*, which are associated with HDL-C production via a system different from RCT, were also considered among the seven major SNPs.<sup>10,12</sup> The genetic variants of *CETP* were observed to exhibit the most significant influence on HDL-C levels, which was concordant with findings from previous reports.<sup>8–10</sup>

Cigarette smoking is associated with lower HDL-C levels, even though the mechanisms are yet to be completely elucidated. Certain studies have shown that smoking is related to ApoA1 concentration<sup>13</sup> and CETP activity<sup>14</sup>; however, these results could

**Table 4.** Interaction between HDL-C levels according to different smoking and drinking statuses and the 7 selected SNPs

	Smoking				<i>P</i> -value for interaction <sup>a</sup>	Drinking				<i>P</i> -value for interaction <sup>a</sup>
	Never & former		Current			Non-moderate		Heavy <sup>b</sup>		
	RR & RA	AA	RR & RA	AA		RR & RA	AA	RR & RA	AA	
	HDL-C (mg/dL)	HDL-C (mg/dL)	HDL-C (mg/dL)	HDL-C (mg/dL)		HDL-C (mg/dL)	HDL-C (mg/dL)	HDL-C (mg/dL)	HDL-C (mg/dL)	
rs3764261	64.0 (63.7–64.3) <i>N</i> = 8,723	72.2 (70.6–73.8) <i>N</i> = 408	56.0 (55.4–56.6) <i>N</i> = 2,130	64.7 (60.7–68.8) <i>N</i> = 109	0.156	62.5 (62.1–62.8) <i>N</i> = 8,464	69.5 (67.9–71.2) <i>N</i> = 405	62.2 (61.6–62.9) <i>N</i> = 2,389	74.6 (70.8–78.4) <i>N</i> = 112	0.015
rs662799	62.9 (62.4–63.3) <i>N</i> = 5,301	66.5 (66.0–67.0) <i>N</i> = 3,830	54.7 (53.9–55.5) <i>N</i> = 1,236	58.5 (57.5–59.5) <i>N</i> = 1,003	0.706	61.5 (61.0–61.9) <i>N</i> = 5,094	64.6 (64.1–65.1) <i>N</i> = 3,775	60.7 (59.9–61.6) <i>N</i> = 1,443	65.6 (64.5–66.6) <i>N</i> = 1,058	0.004
rs1800588	63.6 (63.2–64.0) <i>N</i> = 6,723	66.6 (65.9–67.2) <i>N</i> = 2,408	55.3 (54.6–56.0) <i>N</i> = 1,661	59.5 (58.2–60.8) <i>N</i> = 578	0.312	62.0 (61.6–62.4) <i>N</i> = 6,530	65.0 (64.4–65.7) <i>N</i> = 2,339	61.7 (61.0–62.5) <i>N</i> = 1,854	65.8 (64.5–67.2) <i>N</i> = 647	0.387
rs328	64.3 (63.9–64.6) <i>N</i> = 8,983	70.4 (67.6–73.2) <i>N</i> = 148	56.4 (55.7–57.0) <i>N</i> = 2,207	58.4 (52.3–64.4) <i>N</i> = 32	0.735	62.7 (62.4–63.0) <i>N</i> = 8,721	69.0 (66.1–72.0) <i>N</i> = 148	62.8 (62.1–63.4) <i>N</i> = 2,469	64.8 (59.6–70.1) <i>N</i> = 32	0.658
rs2575876	64.6 (64.2–64.9) <i>N</i> = 8,436	62.1 (61.0–63.3) <i>N</i> = 695	56.6 (55.9–57.2) <i>N</i> = 2,062	54.6 (52.4–56.7) <i>N</i> = 177	0.476	63.0 (62.6–63.3) <i>N</i> = 8,207	60.5 (59.3–61.6) <i>N</i> = 662	63.0 (62.3–63.6) <i>N</i> = 2,291	61.0 (58.6–63.4) <i>N</i> = 210	0.354
rs3786247	63.9 (63.6–64.3) <i>N</i> = 7,232	66.0 (65.3–66.7) <i>N</i> = 1,899	55.6 (55.0–56.3) <i>N</i> = 1,770	59.3 (57.8–60.7) <i>N</i> = 469	0.670	62.3 (62.0–62.7) <i>N</i> = 7,029	64.6 (63.8–65.3) <i>N</i> = 1,840	62.2 (61.5–62.9) <i>N</i> = 1,973	65.0 (63.5–66.5) <i>N</i> = 528	0.569
rs429358	64.4 (64.1–64.7) <i>N</i> = 9,037	61.6 (58.1–65.0) <i>N</i> = 94	56.4 (55.8–57.0) <i>N</i> = 2,214	57.9 (50.1–65.6) <i>N</i> = 25	0.931	62.8 (62.5–63.2) <i>N</i> = 8,773	60.7 (57.4–64.0) <i>N</i> = 96	62.8 (62.1–63.5) <i>N</i> = 2,478	61.1 (52.3–70.0) <i>N</i> = 23	0.723

AA, alternative homo-genotype; BMI, body mass index; HDL-C, high density lipoprotein cholesterol; RA, referent and alternative allele hetero-genotype; RR, referent allele homo-genotype; SNP, single nucleotide polymorphism.

<sup>a</sup>Adjusted for age, sex, smoking, drinking, daily activity, habitual exercise, egg intake, and BMI.

<sup>b</sup>≥20 g alcohol/day.

**Table 5.** Population attributable fractions of non-genetic and genetic factors for low HDL-C

	Proportion of exposure in low HDL-C subjects (%)	OR <sup>a</sup>	95% CI	PAF (%)
Non-genetic factors				
Smoking habit (current)	41.8	2.23	1.85–2.70	23.1
Drinking habit (<20 grams alcohol/day)	76.8	2.19	1.77–2.71	41.8
Daily activity (<8.25 METs/day)	62.7	1.11	0.93–1.33	—
Habitual exercise (<0.73 METs/day)	52.4	1.14	0.95–1.36	—
Egg intake (<3 times/week)	70.4	0.93	0.77–1.12	—
BMI (≥23.0 kg/m <sup>2</sup> )	72.6	2.35	1.93–2.85	41.6
Age (≥57 years)	54.5	1.44	1.20–1.72	16.6
Sex (men)	80.4	4.68	3.72–5.89	63.2
Genetic factors				
rs3764261 (RR)	73.5	1.75	1.44–2.13	31.5
rs662799 (RR)	26.6	2.89	2.35–3.55	17.4
rs1800588 (RR)	27.3	1.16	0.95–1.41	—
rs328 (RR)	81.1	1.36	1.09–1.70	21.6
rs2575876 (RA & AA)	55.6	1.43	1.20–1.71	16.8
rs3786247 (RR)	34.5	1.36	1.13–1.64	9.2
rs429358 (RA & AA)	24.6	1.56	1.27–1.92	8.9
Number of SNPs with high-risk genotype <sup>b</sup>				
0–1 SNPs	7.3	1.00	—	—
2 SNPs	26.4	1.97	1.38–2.82	13.0
3 SNPs	37.4	3.16	2.24–4.47	25.5
4–6 SNPs	29.0	5.49	3.84–7.84	23.7
<i>P</i> for trend			<0.001	

AA, alternative allele homo-genotype; BMI, body mass index; CI, confidence interval; HDL-C, high density lipoprotein cholesterol; OR, odds ratio; PAF, population attributable fraction; RA, referent and alternative allele hetero-genotype; RR, referent allele homo-genotype; SNPs, single nucleotide polymorphisms.

<sup>a</sup>Adjusted for age, sex, smoking, drinking, daily activity, habitual exercise, egg intake and BMI.

<sup>b</sup>SNP of rs1800588 is excluded.

be considered controversial.<sup>32,33</sup> Alcohol consumption is reported to be associated with increased expression of ABCA1<sup>34</sup> and a higher APOA1 concentration<sup>35</sup> in peripheral blood and a lower CETP activity.<sup>36</sup>

In the present study, the interaction of the 65 and seven SNPs with drinking was not statistically significant after Bonferroni correction was applied. Previous studies reported significant association of alcohol consumption and polymorphisms in multiple genes (*CETP*, *APOA1/A2*, *LPL*, *ADH3*, *ADH1*, and *ALDH2*) with HDL-C levels.<sup>37–41</sup> The association between CETP and ABCA1 expressions, and alcohol consumption has been also reported in previous studies, but their mechanism is not clear.<sup>34,36</sup> However, no genome-wide significance was reported in the gene-alcohol interaction for *CETP*, *APOA5*, *LIPC*, and *LPL* in a particular GWAS.<sup>42</sup> The interaction between each SNP and smoking was also not statistically significant after Bonferroni correction was applied. These results suggest that genetic factors may have a minor or negligible impact on the interaction with drinking and smoking.

Several studies have previously reported the association between SNPs and HDL-C levels, which have been listed in the GWAS catalog. In the present study, we selected the 498 SNPs listed in the GWAS results that were a part of the J-MICC Study and observed 65 SNPs with genome-wide significance for the analysis. We selected seven SNPs according to the gene and cytoBand groups. The Manhattan plot for total SNPs consistently showed seven peaks, except that for *SCARB1*. These observations support proposition that the seven SNPs are appropriate representatives of the SNPs associated with HDL-C levels in the present analysis.

In the present study, we investigated the population-based impact of both non-genetic and genetic factors on low HDL-C, using PAF. The OR for low HDL-C was used as the relative risk when the PAF was calculated, because the prevalence of low HDL-C was obtained from the baseline general population and its rate was relatively low (5.0% in both sexes).<sup>30,31</sup> To the best of our knowledge, studies investigating the PAF for low HDL-C with non-genetic and/or genomic factors have not yet been conducted. The highest PAFs was observed in men on the non-genetic factors and in *CETP* rs3764261 on the genetic factors. The impact of the genetic factor PAF was higher than that of smoking and was lower than that of drinking. These observations suggest that, from a public health perspective, the population-based impact of genomic factors for low HDL-C is comparably high compared to non-genetic factors.

The strength of this study is that the population-based impact of non-genetic and genetic factors on HDL-C levels was evaluated simultaneously using data from an adequate number of subjects and total gene information. To our knowledge, this is the first comprehensive report on the population-based impact of the abovementioned factors.

Meanwhile, the present study has several limitations. First, a causal relationship was not confirmed, as this is a cross-sectional study. Second, atheroprotective and non-atheroprotective HDL particles were jointly considered as total HDL-C. The two fractions of HDL2-C and HDL3-C have different effects on CVD risk.<sup>2</sup> Third, the present study selected seven representative SNPs to estimate the population-based impact; the highest impact may have been estimated because the highest coefficients of the seven representative SNPs were selected based on the gene and cytoBand groups. Fourth, the replication test on GWAS was not

conducted, because the present study used information from the GWAS catalog in which the association between SNPs and HDL-C levels had been estimated and published previously. Fifth, the effect of residual SNPs (those apart from the 65 SNPs), referred to as “missing heritability”, was not considered. The polygenic risk score may support the estimation of this effect.<sup>43</sup> Sixth, PAF valid only in the absence of confounding and/or effect modification.<sup>30</sup> The lack of unknown data on confounding is likely to misestimate the true PAF, the extent to which is dependent on the magnitude of confounding.<sup>31</sup> Furthermore, PAF estimate is restricted by time and population and depends on the quality and representativeness of the exposure and risk data.

In conclusion, the present study demonstrated that the population-based impact of genomic factor *CETP* rs3764261 for low HDL-C was higher than that of smoking and lower than that of drinking.

## ACKNOWLEDGMENTS

We thank Dr Nobuyuki Hamajima and Dr Hideo Tanaka for initiating and organizing the J-MICC Study as former principal investigators. We also thank the study participants and the members of the J-MICC Study Group. This study was supported by a Grants-in-Aid for Scientific Research for Priority Areas of Cancer (No. 17015018) and Innovative Areas (No. 221S0001) and by the Japan Society for the Promotion of Science (JSPS) KAKENHI Grant (No. 16H06277 [CoBiA]) from the Japanese Ministry of Education, Culture, Sports, Science and Technology. This study was also supported in part by funds for the BioBank Japan Project received from the Japan Agency for Medical Research and Development since April 2015, and from the Ministry of Education, Culture, Sports, Science and Technology from April 2003 to March 2015.

Conflicts of interest: Dr Nakatochi reports grants from Boehringer Ingelheim outside the submitted work.

## SUPPLEMENTARY MATERIAL

Supplementary data related to this article can be found at <https://doi.org/10.2188/jea.JE20210142>.

## REFERENCES

1. National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). Third Report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III) final report. *Circulation*. 2002;106(25):3143–3421.
2. von Eckardstein A, Kardassis D, editors. *High Density Lipoproteins: From Biological Understanding to Clinical Exploitation*. London: Springer International Publishing; 2015:224.
3. Volcik K, Ballantyne CM, Pownall HJ, Sharrett AR, Boerwinkle E. Interaction effects of high-density lipoprotein metabolism gene variation and alcohol consumption on coronary heart disease risk: the atherosclerosis risk in communities study. *J Stud Alcohol Drugs*. 2007;68(4):485–492.
4. Wada M, Daimon M, Emi M, et al. Genetic association between aldehyde dehydrogenase 2 (ALDH2) variation and high-density lipoprotein cholesterol (HDL-C) among non-drinkers in two large population samples in Japan. *J Atheroscler Thromb*. 2008;15(4):179–184.

5. Chelland Campbell S, Moffatt RJ, Stamford BA. Smoking and smoking cessation — the relationship between cardiovascular disease and lipoprotein metabolism: a review. *Atherosclerosis*. 2008;201(2):225–235.
6. He BM, Zhao SP, Peng ZY. Effects of cigarette smoking on HDL quantity and function: implications for atherosclerosis. *J Cell Biochem*. 2013;114(11):2431–2436.
7. Weissglas-Volkov D, Pajukanta P. Genetic causes of high and low serum HDL-cholesterol. *J Lipid Res*. 2010;51(8):2032–2057.
8. Hiura Y, Shen CS, Kokubo Y, et al. Identification of genetic markers associated with high-density lipoprotein-cholesterol by genome-wide screening in a Japanese population: the Suita study. *Circ J*. 2009;73(6):1119–1126.
9. Nakamura A, Niimura H, Kuwabara K, et al. Gene-gene combination effect and interactions among ABCA1, APOA1, SR-B1, and CETP polymorphisms for serum high-density lipoprotein-cholesterol in the Japanese population. *PLoS One*. 2013;8(12):e82046.
10. Heid IM, Boes E, Müller M, et al. Genome-wide association analysis of high-density lipoprotein cholesterol in the population-based KORA study sheds new light on intergenic regions. *Circ Cardiovasc Genet*. 2008;1(1):10–20.
11. Thompson JF, Wood LS, Pickering EH, Dechairo B, Hyde CL. High-density genotyping and functional SNP localization in the CETP gene. *J Lipid Res*. 2007;48(2):434–443.
12. Surakka I, Horikoshi M, Mägi R, et al; ENGAGE Consortium. The impact of low-frequency and rare variants on lipid levels. *Nat Genet*. 2015;47(6):589–597.
13. Sigurdsson G Jr, Gudnason V, Sigurdsson G, Humphries SE. Interaction between a polymorphism of the apo A-I promoter region and smoking determines plasma levels of HDL and apo A-I. *Arterioscler Thromb*. 1992;12(9):1017–1022.
14. Dullaart RP, Hoogenberg K, Dikkeschei BD, van Tol A. Higher plasma lipid transfer protein activities and unfavorable lipoprotein changes in cigarette-smoking men. *Arterioscler Thromb*. 1994;14(10):1581–1585.
15. Craig WY, Palomaki GE, Haddow JE. Cigarette smoking and serum lipid and lipoprotein concentrations: an analysis of published data. *BMJ*. 1989;298(6676):784–788.
16. Hindorf LA, Sethupathy P, Junkins HA, et al. Potential etiologic and functional implications of genome-wide association loci for human diseases and traits. *Proc Natl Acad Sci USA*. 2009;106(23):9362–9367.
17. Welter D, MacArthur J, Morales J, et al. The NHGRI GWAS Catalog, a curated resource of SNP-trait associations. *Nucleic Acids Res*. 2014;42(Database issue):D1001–D1006.
18. Wakai K, Hamajima N, Okada R, et al; J-MICC Study Group. Profile of participants and genotype distributions of 108 polymorphisms in a cross-sectional study of associations of genotypes with lifestyle and clinical factors: a project in the Japan Multi-Institutional Collaborative Cohort (J-MICC) Study. *J Epidemiol*. 2011;21(3):223–235.
19. Hamajima N; J-MICC Study Group. The Japan Multi-Institutional Collaborative Cohort Study (J-MICC Study) to detect gene-environment interactions for cancer. *Asian Pac J Cancer Prev*. 2007;8(2):317–323.
20. Nishida Y, Hachiya T, Hara M, et al; Japan Multi-Institutional Collaborative Cohort Study Group. The interaction between ABCA1 polymorphism and physical activity on the HDL-cholesterol levels in a Japanese population. *J Lipid Res*. 2020;61(1):86–94.
21. Naito M, Eguchi H, Okada R, et al; J-MICC Study Group. Controls for monitoring the deterioration of stored blood samples in the Japan Multi-Institutional Collaborative Cohort Study (J-MICC Study). *Nagoya J Med Sci*. 2008;70(3–4):107–115.
22. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. *Gigascience*. 2015;4:7.
23. Yamaguchi-Kabata Y, Nakazono K, Takahashi A, et al. Japanese population structure, based on SNP genotypes from 7003 individuals compared to other ethnic groups: effects on population-based association studies. *Am J Hum Genet*. 2008;83(4):445–456.
24. Patterson N, Price AL, Reich D. Population structure and eigenanalysis. *PLoS Genet*. 2006;2(12):e190.
25. 1000 Genomes Project Consortium, Auton A, Brooks LD, et al. A global reference for human genetic variation. *Nature*. 2015;526(7571):68–74.
26. Delaneau O, Zagury JF, Marchini J. Improved whole-chromosome phasing for disease and population genetic studies. *Nat Methods*. 2013;10(1):5–6.
27. Das S, Forer L, Schönherr S, et al. Next-generation genotype imputation service and methods. *Nat Genet*. 2016;48(10):1284–1287.
28. Japanese Ministry of Health, Labour and Welfare. A Basic Direction for Comprehensive Implementation of National Health Promotion. <https://www.mhlw.go.jp/file/06-Seisakujouhou-10900000-Kenkoukyoku/0000047330.pdf>; 2021 Accessed 14.03.2021.
29. Glass S, Dwyer GB; American College of Sports Medicine. ACSM's Metabolic Calculations Handbook. Baltimore: Lippincott Williams & Wilkins publishing; 2007.
30. Rockhill B, Newman B, Weinberg C. Use and misuse of population attributable fractions. *Am J Public Health*. 1998;88(1):15–19.
31. Shield KD, Parkin DM, Whitman DC, et al. Population attributable and preventable fractions: cancer risk factor surveillance, and cancer policy projection. *Curr Epidemiol Rep*. 2016;3:201–211.
32. Kralova Lesna I, Poledne R, Pagacova L, Stavek P, Pitha J. HDL and apolipoprotein A1 concentrations as markers of cholesterol efflux in middle-aged women: interaction with smoking. *Neuro Endocrinol Lett*. 2012;33(Suppl 2):38–42.
33. Freeman DJ, Caslake MJ, Griffin BA, et al. The effect of smoking on post-heparin lipoprotein and hepatic lipase, cholesteryl ester transfer protein and lecithin:cholesterol acyl transferase activities in human plasma. *Eur J Clin Invest*. 1998;28(7):584–591.
34. Hoang A, Tefft C, Duffy SJ, et al. ABCA1 expression in humans is associated with physical activity and alcohol consumption. *Atherosclerosis*. 2008;197(1):197–203.
35. Rimm EB, Williams P, Fosher K, Criqui M, Stampfer MJ. Moderate alcohol intake and lower risk of coronary heart disease: meta-analysis of effects on lipids and haemostatic factors. *BMJ*. 1999;319(7224):1523–1528.
36. Hannuksela M, Marcel YL, Kesäniemi YA, Savolainen MJ. Reduction in the concentration and activity of plasma cholesteryl ester transfer protein by alcohol. *J Lipid Res*. 1992;33(5):737–744.
37. Mehlhlig K, Strandhagen E, Svensson PA, et al. CETP TaqIB genotype modifies the association between alcohol and coronary heart disease: the INTERGENE case-control study. *Alcohol*. 2014;48(7):695–700.
38. De Oliveira E Silva ER, Foster D, McGee Harper M, et al. Alcohol consumption raises HDL cholesterol levels by increasing the transport rate of apolipoproteins A-I and A-II. *Circulation*. 2000;102(19):2347–2352.
39. Baik I, Lee S, Kim SH, Shin C. A lipoprotein lipase gene polymorphism interacts with consumption of alcohol and unsaturated fat to modulate serum HDL-cholesterol concentrations. *J Nutr*. 2013;143(10):1618–1625.
40. Hines LM, Stampfer MJ, Ma J, et al. Genetic variation in alcohol dehydrogenase and the beneficial effect of moderate alcohol consumption on myocardial infarction. *N Engl J Med*. 2001;344(8):549–555.
41. Yokoyama A, Yokoyama T, Matsui T, et al. Alcohol Dehydrogenase-1B (rs1229984) and Aldehyde Dehydrogenase-2 (rs671) Genotypes Are Strong Determinants of the Serum Triglyceride and Cholesterol Levels of Japanese Alcoholic Men. *PLoS One*. 2015;10(8):e0133460.
42. Marques-Vidal P, Bochud M, Paccaud F, et al. No interaction between alcohol consumption and HDL-related genes on HDL cholesterol levels. *Atherosclerosis*. 2010;211(2):551–557.
43. Warren M. The approach to predictive medicine that is taking genomics research by storm. *Nature*. 2018;562(7726):181–183.